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(54) Title: BINDING DOMAINS FROM PLASMODIUM VIVAX AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING PROTEINS		
(57) Abstract <p>The present invention provides isolated polypeptides useful in the treatment and prevention of malaria caused by <i>Plasmodium falciparum</i> or <i>P. vivax</i>. In particular, the polypeptides are derived from the binding domains of the proteins in the EBL family as well as the sialic acid binding protein (SABP) on <i>P. falciparum</i> merozoites. The polypeptides may also be derived from the Duffy antigen binding protein (DABP) on <i>P. vivax</i> merozoites.</p>		

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BINDING DOMAINS FROM PLASMODIUM VIVAX
AND PLASMODIUM FALCIPARUM ERYTHROCYTE
BINDING PROTEINS

BACKGROUND OF THE INVENTION

Malaria infects 200 - 400 million people each year causing 1-2 million deaths, thus remaining one of the most important infectious diseases in the world. Approximately 25 percent of all deaths of children in rural Africa between the ages of one and four years are caused by malaria. Due to the importance of the disease as a worldwide health problem, considerable effort is being expended to identify and develop malaria vaccines.

Malaria in humans is caused by four species of the parasite *Plasmodium*: *P. falciparum*, *P. vivax*, *P. knowlesi* and *P. malariae*. The major cause of malaria in humans is *P. falciparum* which infects 200 million to 400 million people every year, killing 1 to 4 million.

P. vivax (one of the four species infective to humans) cannot be cultured in vitro, as has been possible with *P. knowlesi* (a malarial strain found in old world monkeys which also invade human erythrocytes) and *P. falciparum*. Although *P. vivax* bears substantial phylogenetic similarity to *P. knowlesi*, the two species are different in many important respects. For example, *P. vivax* is not infective of many simian species and infection is poorly established in others, whereas *P. knowlesi* is poorly infective of humans while readily infecting many simian species.

The basis of various potential vaccines to combat malaria is appreciated through an understanding of the life cycle of the parasite. Infection in humans begins when young malarial parasites or "sporozoites" are injected into the bloodstream of a human by the mosquito. Following injection, the parasite localizes to liver cells. After approximately

one week the parasites or "merozoites" are released into the bloodstream. The entry of the parasites into the bloodstream begins the "erythrocytic" phase. Each parasite enters the red blood cell in order to grow and develop. When the merozoite matures in the red blood cell, it is known as a trophozoite. The trophozoite undergoes several rounds of nuclear division (schizogony) until it ruptures the erythrocyte, releasing from 6 to 24 merozoites. After several asexual schizogonic cycles, some parasites, instead of becoming schizonts through asexual reproduction, develop into morphologically distinct forms known as "gametocytes" which are long-lived and undergo sexual development.

Sexual development of the malaria parasites involve the female or "macrogametocyte" and the male parasite or "microgametocyte." These gametocytes do not undergo any further development in humans. Upon ingestion of the gametocytes into the mosquito, the complicated sexual cycle begins in the midgut of the mosquito. The red blood cells disintegrate in the midgut of the mosquito after 10 to 20 minutes. The microgametocyte continues to develop through exflagellation and releases 8 highly flagellated microgametes. Fertilization occurs upon fusion of the microgamete and the macrogamete. The fertilized parasite is known as a zygote which develops into an "ookinete." The ookinete embeds in the midgut of the mosquito, transforming into an oocyst within which many small sporozoites form. Before embedding in the midgut, the ookinete must first penetrate the peritrophic membrane which apparently acts as a barrier for invasion of ingested parasites. When the oocyst ruptures the sporozoites migrate to the salivary gland of the mosquito via the hemolymph. Once in the saliva of the mosquito, the parasite can be injected into a host.

The erythrocytic stage of the *Plasmodium* life cycle is of special relevance to vaccine development because the clinical and pathologic features of malaria in the host are attributable to this stage. In *P. vivax*, and *P. knowlesi*, Duffy blood group determinants present on Duffy positive erythrocytes are essential for invasion of human erythrocytes

(Miller et al., Science 189: 561-563, (1975); Miller et al., N. Engl. J. Med. 295: 302-304, (1976)). In *P. falciparum*, invasion of merozoites into erythrocytes appears to be dependent on binding to sialic acids on glycoporphins on the erythrocyte (Miller, et al., J. Exp. Med. 146: 277-281, (1971); Pasvol, et al., Lancet. ii: 947-950 (1982); Pasvol, et al., Nature, 279: 64-66 (1982); Perkins, J. Exp. Med. 160: 788-798 (1984)). Studies with the monkey parasite *P. knowlesi* allow a clearer understanding of the multiple events that occur during invasion. It is likely that even though *P. vivax* and *P. falciparum* bind to the Duffy antigen and sialic acids respectively, they share common strategies of invasion with each other and with *P. knowlesi*.

In *P. knowlesi*, during invasion a merozoite first attaches to an erythrocyte on any surface of the merozoite, then reorients so that its apical end is in contact with the erythrocyte (Dvorak et al., Science 187: 748-750, (1975)). Both attachment and reorientation of merozoites occur equally well on Duffy positive and Duffy negative cells. A junction then forms between the apical end of the merozoite and the Duffy positive erythrocyte followed by vacuole formation and entry of the merozoite into the vacuole. Aikawa et al., J. Cell Biol. 77: 72-82 (1978). Junction formation and merozoite entry into the erythrocyte do not occur on Duffy negative cells (Miller et al., J. Exp. Med. 149: 172-184 (1979)), suggesting that a receptor specific for the Duffy determinant is involved in apical junction formation but not initial attachment.

The apical end of the merozoite is defined by the presence of three organelles: rhoptries, dense granules and micronemes. The rhoptries and dense granules release their contents at vacuole formation (Ladda et al., 1969; Aikawa et al., J. Cell Biol., 77: 72-82 (1978); Torn et al., Infection and Immunity 57: 3230-3233 (1989); Bannister and Dluzewski, Blood Cells 16: 257-292 (1990)). To date the function of the microneme is unknown. Nevertheless, the location of the micronemes suggest that they are involved in the invasion process. Duffy Antigen Binding Protein (DABP) and Sialic Acid

Binding Protein (SABP) have been localized to the micronemes of *P. knowlesi* and *P. falciparum* respectively (Adams et al., Cell 63: 141-153 (1990); Sim et al., Mol. Biochem. Parasitol. 51: 157-160 (1992)).

5 DABP and SABP are soluble proteins that appear in the culture supernatant after infected erythrocytes release merozoites. Immunochemical data indicate that DABP and SABP which are the respective ligands for the *P. vivax* and *P. falciparum* Duffy and sialic acid receptors on erythrocytes, 10 possess specificities of binding which are identical either in soluble or membrane bound form.

DABP is a 135 kDa protein which binds specifically to Duffy blood group determinants (Wertheimer et al., Exp. Parasitol. 69: 340-350 (1989); Barnwell, et al., J. Exp. Med. 15 169: 1795-1802 (1989)). Thus, binding of DABP is specific to human Duffy positive erythrocytes. There are four major Duffy phenotypes for human erythrocytes: Fy(a), Fy(b), Fy(ab) and Fy(negative), as defined by the anti-Fy^a and anti-Fy^b sera (Hadley et al., In Red Cell Antigens and Antibodies, G. 20 Garratty, ed. (Arlington, Va.:American Association of Blood Banks) pp. 17-33 (1986)). DABP binds equally to both Fy(a) and Fy(b) erythrocytes which are equally susceptible to invasion by *P. vivax*; but not to Fy(negative) erythrocytes.

In the case of SABP, a 175kDa protein, binding is 25 specific to the glycophorin sialic acid residues on erythrocytes (Camus and Hadley, Science 230:553-556 (1985); Orlandi, et al., J. Cell Biol. 116:901-909 (1992)). Thus, neuraminidase treatment (which cleaves off sialic acid residues) render erythrocytes immune to *P. falciparum* 30 invasion.

The specificities of binding and correlation to invasion by the parasite thus indicate that DABP and SABP are the proteins of *P. vivax* and *P. falciparum* which interact with sialic acids and the Duffy antigen on the erythrocyte. The 35 genes encoding both proteins have been cloned and the DNA and predicted protein sequences have been determined (B. Kim Lee Sim, et al., J. Cell Biol. 111: 1877-1884 (1990); Fang, X., et al., Mol. Biochem Parasitol. 44: 125-132 (1991)).

Despite considerable research efforts worldwide, because of the complexity of the *Plasmodium* parasite and its interaction with its host, it has not been possible to discover a satisfactory solution for prevention or abatement of the blood stage of malaria. Because malaria is a such a large worldwide health problem, there is a need for methods that abate the impact of this disease. The present invention provides effective preventive and therapeutic measures against *Plasmodium* invasion.

SUMMARY OF THE INVENTION

The present invention provides compositions comprising an isolated DABP binding domain polypeptides and/or isolated SABP binding domain polypeptides. The DABP binding domain polypeptides preferably comprise between about 200 and about 300 amino acid residues while the SABP binding domain polypeptides preferably comprises between about 200 and about 600 amino acid residues. A preferred DABP binding domain polypeptide has residues 1 to about 325 of the amino acid sequence found in SEQ ID No. 2. A preferred SABP binding domain polypeptide has residues 1 to about 616 of the amino acid sequence of SEQ ID No. 4.

The present invention also includes pharmaceutical compositions comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* merozoites in an organism. In addition, isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* may be added to the pharmaceutical composition.

Also provided are pharmaceutical compositions comprising a pharmaceutically acceptable carrier and an isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* merozoites in an organism. In addition, isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to

Plasmodium vivax may be added to the pharmaceutical composition.

Isolated polynucleotides which encode a DABP binding domain polypeptides or SABP binding domain polypeptides are also disclosed. In addition, the present invention includes a recombinant cell comprising the polynucleotide encoding the DABP binding domain polypeptide.

The current invention further includes methods of inducing a protective immune response to *Plasmodium* merozoites in a patient. The methods comprise administering to the patient an immunologically effective amount of a pharmaceutical composition comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide, an SABP binding domain polypeptide or a combination thereof.

The present disclosure also provides DNA sequences from additional *P. falciparum* genes in the erythrocyte binding ligand (EBL) family that have regions conserved with the *P. falciparum* 175 kD and *P. vivax* 135 kD binding proteins.

DEFINITIONS

As used herein a "DABP binding domain polypeptide" or a "SABP binding domain polypeptide" are polypeptides substantially identical (as defined below) to a sequence from the cysteine-rich, amino-terminal region of the Duffy antigen binding protein (DABP) or sialic acid binding protein (SABP), respectively. Such polypeptides are capable of binding either the Duffy antigen or sialic acid residues on glycophorin. In particular, DABP binding domain polypeptides consist of amino acid residues substantially similar to a sequence of SABP within a binding domain from the N-terminal amino acid (residue 1) to about residue 325. SABP binding domain polypeptides consist of residues substantially similar to a sequence of DABP within a binding domain from the N-terminal amino acid (residue 1) to about residue 616.

The binding domain polypeptides encoded by the genes of the EBL family consist of those residues substantially identical to the sequence of the binding domains of DABP and

SABP as defined above. The EBL family comprises sequences with substantial similarity to the conserved regions of the DABP and SABP. These include those sequences reported here as EBL-e1 (SEQ ID NOs 5 and 6), E31a (SEQ ID NOs 7 and 8), EBL-e2 (SEQ ID NOs 9 and 10) and Proj3 (SEQ ID NOs 11 and 12).

The polypeptides of the invention can consist of the full length binding domain or a fragment thereof. Typically DABP binding domain polypeptides will consist of from about 50 to about 325 residues, preferably between about 75 and 300, more preferably between about 100 and about 250 residues. SABP binding domain polypeptides will consist of from about 50 to about 616 residues, preferably between about 75 and 300, more preferably between about 100 and about 250 residues.

Particularly preferred polypeptides of the invention are those within the binding domain that are conserved between SABP and the EBL family. Residues within these conserved domains are shown in Figure 1, below.

Two polynucleotides or polypeptides are said to be "identical" if the sequence of nucleotides or amino acid residues in the two sequences is the same when aligned for maximum correspondence. Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman *Adv. Appl. Math.* 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson and Lipman *Proc. Natl. Acad. Sci. (U.S.A.)* 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by inspection. These references are incorporated herein by reference.

The term "substantial identity" means that a polypeptide comprises a sequence that has at least 80% sequence identity, preferably 90%, more preferably 95% or more, compared to a reference sequence over a comparison window of about 20 residues to about 600 residues-- typically about 50 to about 500 residues usually about 250 to 300 residues. The values of percent identity are determined using

the programs above. Particularly preferred peptides of the present invention comprise a sequence in which at least 70% of the cysteine residues conserved in DABP and SABP are present. Additionally, the peptide will comprise a sequence in which at least 50% of the Tryptophan residues conserved in DABP and SABP are present. The term substantial similarity is also specifically defined here with respect to those amino acid residues found to be conserved between DABP, SABP and the sequences of the EBL family. These conserved amino acids consist prominently of tryptophan and cysteine residues conserved among all sequences reported here. In addition the conserved amino acid residues include phenylalanine residues which may be substituted with tyrosine. These amino acid residues may be determined to be conserved after the sequences have been aligned using methods outlined above by someone skilled in the art.

Another indication that polypeptide sequences are substantially identical is if one protein is immunologically reactive with antibodies raised against the other protein. Thus, the polypeptides of the invention include polypeptides immunologically reactive with antibodies raised against the SABP binding domain, the DABP binding domain or raised against the conserved regions of the EBL family.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. Stringent conditions are sequence dependent and will be different in different circumstances. Generally, stringent conditions are selected to be about 5° C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typically, stringent conditions will be those in which the salt concentration is at least about 0.02 molar at pH 7 and the temperature is at least about 60°C.

The phrases "isolated" or "biologically pure" refer to material which is substantially or essentially free from components which normally accompany it as found in its native

state. Thus, the binding domain polypeptides of this invention do not contain materials normally associated with their *in situ* environment, e.g., other proteins from a merozoite membrane. However, even where a protein has been isolated to a homogenous or dominant band by PAGE, there can be trace contaminants in the range of 5-10% of native protein which co-purify with the desired protein. Isolated polypeptides of this invention do not contain such endogenous co-purified protein.

Protein purity or homogeneity may be indicated by a number of means well known in the art, such as polyacrylamide gel electrophoresis of a protein sample, followed by visualization upon staining. For certain purposes high resolution will be needed and HPLC or a similar means for purification utilized.

The term "residue" refers to an amino acid (D or L) or amino acid mimetic incorporated in a oligopeptide by an amide bond or amide bond mimetic. An amide bond mimetic of the invention includes peptide backbone modifications well known to those skilled in the art.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 represents an alignment of the predicted amino acid sequences of the DABP binding domain (Vivax), the two homologous SABP domains (SABP F1 and SABP F2) and the sequenced members of the EBL gene family (ebl-e1, E31a, EBL-e2 and the three homologous Proj3 domains.

Figure 2 represents a schematic of the pRE4 cloning vector.

Figure 3 shows primers useful for isolating sequences encoding the conserved motifs of the invention.

DESCRIPTION OF THE PREFERRED EMBODIMENT

The binding of merozoites and schizonts to erythrocytes is mediated by specific binding proteins on the surface of the merozoite or schizont and is necessary for erythrocyte invasion. In the case of *P. falciparum*, this binding involves specific interaction between sialic acid

glycophorin residues on the erythrocyte and the sialic acid binding protein (SABP) on the surface of the merozoite or schizont. The ability of purified SABP to bind erythrocytes with chemically or enzymatically altered sialic acid residues paralleled the ability of *P. falciparum* to invade these erythrocytes. Furthermore, sialic acid deficient erythrocytes neither bind SABP nor support invasion by *P. falciparum*. The DNA encoding SABP from *P. falciparum* has also been cloned and sequenced.

In *P. vivax*, specific binding to the erythrocytes involves interaction between the Duffy blood group antigen on the erythrocyte and the Duffy antigen binding protein (DABP) on the merozoite. Duffy binding proteins were defined biologically as those soluble proteins that appear in the culture supernatant after the infected erythrocytes release merozoites which bind to human Duffy positive, but not to human Duffy negative erythrocytes. It has been shown that binding of the *P. vivax* DABP protein to Duffy positive erythrocytes is blocked by antisera to the Duffy blood group determinants. Purified Duffy blood group antigens also block the binding to erythrocytes. DABP has also been shown to bind Duffy blood group determinants on Western blots.

Duffy positive blood group determinants on human erythrocytes are essential for invasion of human erythrocytes by *Plasmodium vivax*. Both attachment and reorientation of *P. vivax* merozoites occur equally well on Duffy positive and negative erythrocytes. A junction then forms between the apical end of the merozoite and the Duffy-positive erythrocyte, followed by vacuole formation and entry of the merozoite into the vacuole. Junction formation and merozoite entry into the erythrocyte do not occur on Duffy negative cells, suggesting that the receptor specific for the Duffy determinant is involved in apical junction formation but not initial attachment. The DNA sequences encoding the DABP from *P. vivax* and *P. knowlesi* have been cloned and sequenced.

P. vivax red cell invasion has an absolute requirement for the Duffy blood group antigen. Isolates of *P. falciparum*, however, vary in their dependency on sialic acid

for invasion. Certain *P. falciparum* clones have been developed which invade sialic acid deficient erythrocytes at normal rates. This suggests that certain strains of *P. falciparum* can interact with other ligands on the erythrocyte and so may possess multiple erythrocyte binding proteins with differing specificities.

A basis for the present invention is the discovery of the binding domains in both DABP and SABP. Comparison of the predicted protein sequences of DABP and SABP reveals an amino-terminal, cysteine-rich region in both proteins with a high degree of similarity between the two proteins. The amino-terminal, cysteine-rich region of DABP contains about 325 amino acids, whereas the amino-terminal, cysteine-rich region of SABP contains about 616 amino acids. This is due to an apparent duplication of the amino-terminal, cysteine-rich region in the SABP protein. The cysteine residues are conserved between the two regions of SABP and DABP, as are the amino acids surrounding the cysteine residues and a number of aromatic amino acid residues in this region. The amino-terminal cysteine rich region and another cysteine-rich region near the carboxyl-terminus show the most similarity between the DABP and SABP proteins. The region of the amino acid sequence between these two cysteine-rich regions show only limited similarity between DABP and SABP.

Other *P. falciparum* open reading frames and genes with regions that have substantial identity to binding domains of SABP and DABP have been identified. Multiple copies of these sequences exist in the parasite genome, indicating their important activity in host-parasite interactions. A family of these sequences (the EBL family) have been cloned from chromosome 7 subsegment libraries that were constructed during genetic studies of the chloroquine resistance locus (Wellems et. al., PNAS 88: 3382-3386 (1991)). Alignment of EBL sequences identified domains highly conserved with the *P. falciparum* 175 kD protein; these conserved domains have in turn been used to identify genes (ebl-e1, ebl-e2) one of which (ebl-e1) resides on chromosome 13. Genetic linkage studies have placed this gene within a region of chromosome 13 that

affects invasion of malarial parasites in human red blood cells (Wellems et al., Cell 49:633-642 (1987)).

Southern hybridization experiments using probes from these open reading frames have indicated that additional copies of these conserved sequences are located elsewhere in the genome. The largest of the open reading frames on chromosome 7 is 8 kilobases and contains four tandem repeats homologous to the N-terminal, cysteine-rich unit of SABP and DABP.

Figure 1 represents an alignment of the EBL family with the DABP binding domain and two homologous regions of SABP (F₁ and F₂). The EBL family is divided into two sub-families to achieve optimal alignment. Conserved cysteine residues are shown in bold face and conserved aromatic residues are underlined.

The polypeptides of the invention can be used to raise monoclonal antibodies specific for the binding domains of SABP, DABP or the conserved regions in the EBL gene family. The antibodies can be used for diagnosis of malarial infection or as therapeutic agents to inhibit binding of merozoites to erythrocytes. The production of monoclonal antibodies against a desired antigen is well known to those of skill in the art and is not reviewed in detail here.

The multitude of techniques available to those skilled in the art for production and manipulation of various immunoglobulin molecules can thus be readily applied to inhibit binding. As used herein, the terms "immunoglobulin" and "antibody" refer to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. Immunoglobulins may exist in a variety of forms besides antibodies, including for example, Fv, Fab, and F(ab)₂, as well as in single chains. For a general review of immunoglobulin structure and function see, *Fundamental Immunology*, 2d Ed., W.E. Paul ed., Ravens Press, N.Y., (1989).

Antibodies which bind polypeptides of the invention may be produced by a variety of means. The production of non-human monoclonal antibodies, e.g., murine, lagomorpha, equine, etc., is well known and may be accomplished by, for

example, immunizing the animal with a preparation containing the polypeptide. Antibody-producing cells obtained from the immunized animals are immortalized and screened, or screened first for the production of antibody which inhibits binding between merozoites and erythrocytes and then immortalized. For a discussion of general procedures of monoclonal antibody production see Harlow and Lane, *Antibodies, A Laboratory Manual* Cold Spring Harbor Publications, N.Y. (1988).

Thus, the present invention allows targeting of protective immune responses or monoclonal antibodies to sequences in the binding domains that are conserved between SABP, DABP and encoded regions of the EBL family. Identification of the binding regions of these proteins facilitates vaccine development because it allows for a focus of effort upon the functional elements of the large molecules. The particular sequences within the binding regions refine the target to critical regions that have been conserved during evolution, and are thus preferred for use as vaccines against the parasite.

The genes of the EBL family (which have not previously been sequenced) can be used as markers to detect the presence of the *P. falciparum* parasite in patients. This can be accomplished by means well known to practitioners in the art using tissue or blood from symptomatic patients in PCR reactions with oligonucleotides complementary to portions of the genes of the EBL family. Furthermore, sequencing the EBL family provides a means for skilled practitioners to generate defined probes to be used as genetic markers in a variety of applications.

Additionally, the present invention defines a conserved motif present in, but not restricted to other members of the subphylum Apicomplexa which participates in host parasite interaction. This motif can be identified in *Plasmodium* species and other parasitic protozoa by the polymerase chain reaction using the synthetic oligonucleotide primers shown in Figure 3. PCR methods are described in detail below. These primers are designed from regions in the conserved motif showing the highest degree of conservation

among DABP, SABP and the EBL family. Figure 3 shows these regions and the consensus amino acid sequences derived from them.

A. General Methods

Much of the nomenclature and general laboratory procedures required in this application can be found in Sambrook, et al., *Molecular Cloning A Laboratory Manual* (2nd Ed.), Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1989. The manual is hereinafter referred to as "Sambrook, et al."

B. Methods for isolating DNA encoding SABP, DABP and EBL binding regions

The nucleic acid compositions of this invention, whether RNA, cDNA, genomic DNA, or a hybrid of the various combinations, may be isolated from natural sources or may be synthesized in vitro. The nucleic acids claimed may be present in transformed or transfected whole cells, in a transformed or transfected cell lysate, or in a partially purified or substantially pure form.

Techniques for nucleic acid manipulation of genes encoding the binding domains of the invention, such as subcloning nucleic acid sequences encoding polypeptides into expression vectors, labelling probes, DNA hybridization, and the like are described generally in Sambrook et al., incorporated herein by reference.

Recombinant DNA techniques can be used to produce the binding domain polypeptides. In general, the DNA encoding the SABP and DABP binding domains are first cloned or isolated in a form suitable for ligation into an expression vector. After ligation, the vectors containing the DNA fragments or inserts are introduced into a suitable host cell for expression of the recombinant binding domains. The polypeptides are then isolated from the host cells.

There are various methods of isolating the DNA sequences encoding the SABP, DABP and EBL binding domains. Typically, the DNA is isolated from a genomic or cDNA library using labelled oligonucleotide probes specific for sequences in the DNA. Restriction endonuclease digestion of genomic DNA

or cDNA containing the appropriate genes can be used to isolate the DNA encoding the binding domains of these proteins. Since the DNA sequences of the SABP and DABP genes are known, a panel of restriction endonucleases can be constructed to give cleavage of the DNA in the desired regions. After restriction endonuclease digestion, DNA encoding SABP binding domain or DABP binding domain is identified by its ability to hybridize with nucleic acid probes, for example on Southern blots, and these DNA regions are isolated by standard methods familiar to those of skill in the art. See Sambrook, et al.

The polymerase chain reaction can also be used to prepare DABP, SABP EBL binding domain DNA. Polymerase chain reaction technology (PCR) is used to amplify nucleic acid sequences of the DABP and SABP binding domains directly from mRNA, from cDNA, and from genomic libraries or cDNA libraries. The primers shown in Figure 3 are particularly preferred for this process.

Appropriate primers and probes for amplifying the SABP and DABP binding region DNA's are generated from analysis of the DNA sequences. In brief, oligonucleotide primers complementary to the two 3' borders of the DNA region to be amplified are synthesized. The polymerase chain reaction is then carried out using the two primers. See *PCR Protocols: A Guide to Methods and Applications*. (Innis, M, Gelfand, D., Sninsky, J. and White, T., eds.), Academic Press, San Diego (1990). Primers can be selected to amplify the entire DABP regions or to amplify smaller segments of the DABP and SABP binding domains, as desired.

Oligonucleotides for use as probes are chemically synthesized according to the solid phase phosphoramidite triester method first described by Beaucage, S.L. and Caruthers, M.H., 1981, *Tetrahedron Letts.*, 22(20):1859-1862 using an automated synthesizer, as described in Needham-VanDevanter, D.R., et al. 1984, *Nucleic Acids Res.*, 12:6159-6168. Purification of olig nucleotides is by either native acrylamide gel electrophoresis or by anion-exchange

HPLC as described in Pearson, J.D. and Regnier, F.E., 1983, J. Chrom., 255:137-149.

5 The sequence of the synthetic oligonucleotides can be verified using the chemical degradation method of Maxam, A.M. and Gilbert, 1980, in W., Grossman, L. and Moldave, D., eds. Academic Press, New York, Methods in Enzymology, 65:499-560.

10 Other methods known to those of skill in the art may also be used to isolate DNA encoding all or part of the SABP or DABP binding domains. See Sambrook, et al.

C. Expression of DABP, SABP and EBL Binding Domain Polypeptides

15 Once the binding domain DNAs are isolated and cloned, one may express the desired polypeptides in a recombinantly engineered cell such as bacteria, yeast, insect (especially employing baculoviral vectors), and mammalian cells. It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of the DNA encoding the DABP and SABP binding domains. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

20 In brief summary, the expression of natural or synthetic nucleic acids encoding binding domains will typically be achieved by operably linking the DNA or cDNA to a promoter (which is either constitutive or inducible), followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding the binding domains. To obtain high level expression of a cloned gene, it is desirable to construct expression plasmids which contain, at the minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator.

1. Expression in Prokaryotes

Examples of regulatory regions suitable for this purpose in *E. coli* are the promoter and operator region of the *E. coli* tryptophan biosynthetic pathway as described by Yanofsky, C., 1984, *J. Bacteriol.*, 158:1018-1024 and the leftward promoter of phage lambda (P_L) as described by Herskowitz, I. and Hagen, D., 1980, *Ann. Rev. Genet.*, 14:399-445. The inclusion of selection markers in DNA vectors transformed in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol. See Sambrook et al. for details concerning selection markers for use in *E. coli*.

The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA.

Expression systems for expressing the DABP and SABP binding domains are available using *E. coli*, *Bacillus* sp. (Palva, I et al., 1983, *Gene* 22:229-235; Mosbach, K. et al. *Nature*, 302:543-545 and *Salmonella*. *E. coli* systems are preferred.

The binding domain polypeptides produced by prokaryote cells may not necessarily fold properly. During purification from *E. coli*, the expressed polypeptides may first be denatured and then renatured. This can be accomplished by solubilizing the bacterially produced proteins in a chaotropic agent such as guanidine HCl and reducing all the cysteine residues with a reducing agent such as beta-mercaptoethanol. The polypeptides are then renatured, either by slow dialysis or by gel filtration. U.S. Patent No. 4,511,503.

Detection of the expressed antigen is achieved by methods known in the art as radioimmunoassays, Western blotting techniques or immunoprecipitation. Purification from *E. coli* can be achieved following procedures described in U.S. Patent No. 4,511,503.

2. Synthesis of SABP, DABP and EBL Binding Domains in Eukaryotes

A variety of eukaryotic expression systems such as yeast, insect cell lines and mammalian cells, are known to those of skill in the art. As explained briefly below, the DABP and SABP binding domains may also be expressed in these eukaryotic systems.

a. Expression in Yeast

Synthesis of heterologous proteins in yeast is well known and described. *Methods in Yeast Genetics*, Sherman, F., et al., Cold Spring Harbor Laboratory, (1982) is a well recognized work describing the various methods available to produce the binding domains in yeast.

Examples of promoters for use in yeast include GAL1,10 (Johnson, M., and Davies, R.W., 1984, Mol. and Cell. Biol., 4:1440-1448) ADH2 (Russell, D., et al. 1983, J. Biol. Chem., 258:2674-2682), PH05 (EMBO J. 6:675-680, 1982), and MF α 1 (Herskowitz, I. and Oshima, Y., 1982, in *The Molecular Biology of the Yeast Saccharomyces*, (eds. Strathern, J.N. Jones, E.W., and Broach, J.R., Cold Spring Harbor Lab., Cold Spring Harbor, N.Y., pp. 181-209. A multicopy plasmid with a selective marker such as Leu-2, URA-3, Trp-1, and His-3 is also desirable.

A number of yeast expression plasmids like YEp6, YEp13, YEp4 can be used as vectors. A gene of interest can be fused to any of the promoters in various yeast vectors. The above-mentioned plasmids have been fully described in the literature (Botstein, et al., 1979, Gene, 8:17-24; Broach, et al., 1979, Gene, 8:121-133).

Two procedures are used in transforming yeast cells. In one case, yeast cells are first converted into protoplasts using zymolyase, lyticase or glucylase, followed by addition of DNA and polyethylene glycol (PEG). The PEG-treated protoplasts are then regenerated in a 3% agar medium under selective conditions. Details of this procedure are given in the papers by J.D. Beggs, 1978, Nature (London), 275:104-109; and Hinnen, A., et al., 1978, Proc. Natl. Acad. Sci. USA, 75:1929-1933. The second procedure does not involve removal

of the cell wall. Instead the cells are treated with lithium chloride or acetate and PEG and put on selective plates (Ito, H., et al., 1983, J. Bact., 153:163-168).

The binding domains can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassays of other standard immunoassay techniques.

b. Expression in Mammalian and Insect Cell Cultures

Illustrative of cell cultures useful for the production of the binding domains are cells of insect or mammalian origin. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. Illustrative examples of mammalian cell lines include VERO and HeLa cells, Chinese hamster ovary (CHO) cell lines, W138, BHK, Cos-7 or MDCK cell lines.

As indicated above, the vector, e. g., a plasmid, which is used to transform the host cell, preferably contains DNA sequences to initiate transcription and sequences to control the translation of the antigen gene sequence. These sequences are referred to as expression control sequences. When the host cell is of insect or mammalian origin illustrative expression control sequences are obtained from the SV-40 promoter (Science, 222:524-527, 1983), the CMV I.E. Promoter (Proc. Natl. Acad. Sci. 81:659-663, 1984) or the metallothionein promoter (Nature 296:39-42, 1982). The cloning vector containing the expression control sequences is cleaved using restriction enzymes and adjusted in size as necessary or desirable and ligated with DNA coding for the SABP or DABP polypeptides by means well known in the art.

As with yeast, when higher animal host cells are employed, polyadenylation or transcription terminator sequences from known mammalian genes need to be incorporated into the vector. An example of a terminator sequence is the polyadenylation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be

included. An example of a splicing sequence is the VP1 intron from SV40 (Sprague, J. et al., 1983, J. Virol. 45: 773-781).

Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors.

Saveria-Campo, M., 1985, "Bovine Papilloma virus DNA as a Eukaryotic Cloning Vector" in DNA Cloning Vol. II a Practical Approach Ed. D.M. Glover, IRL Press, Arlington, Virginia pp. 213-238.

The host cells are competent or rendered competent for transformation by various means. There are several well-known methods of introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextran, electroporation and micro-injection of the DNA directly into the cells.

The transformed cells are cultured by means well known in the art. Biochemical Methods in Cell Culture and Virology, Kuchler, R.J., Dowden, Hutchinson and Ross, Inc., (1977). The expressed DABP and SABP binding domain polypeptides are isolated from cells grown as suspensions or as monolayers. The latter are recovered by well known mechanical, chemical or enzymatic means.

c. Expression in recombinant vaccinia virus- or adenovirus-infected cells

In addition to use in recombinant expression systems, the isolated binding domain DNA sequences can also be used to transform viruses that transfect host cells in the patient. Live attenuated viruses, such as vaccinia or adenovirus, are convenient alternatives for vaccines because they are inexpensive to produce and are easily transported and administered. Vaccinia vectors and methods useful in immunization protocols are described, for example, in U.S. Patent No. 4,722,848, incorporated herein by reference.

Suitable viruses for use in the present invention include, but are not limited to, pox viruses, such as canarypox and cowpox viruses, and vaccinia viruses, alpha

viruses, adenoviruses, and other animal viruses. Th
recombinant viruses can be produced by methods well known in
th art, for example, using homologous recombination r
ligating two plasmids. A recombinant canarypox or cowpox
5 virus can be made, for example, by inserting the DNA's
encoding the DABP and SABP binding domain polypeptides into
plasmids so that they are flanked by viral sequences on both
sides. The DNA's encoding the binding domains are then
inserted into the virus genome through homologous
10 recombination.

A recombinant adenovirus can be produced, for
example, by ligating together two plasmids each containing
about 50% of the viral sequence and the DNA sequence encoding
erythrocyte binding domain polypeptide. Recombinant RNA
15 viruses such as the alpha virus can be made via a CDNA
intermediate using methods known in the art.

In the case of vaccinia virus (for example, strain
WR), the DNA sequence encoding the binding domains can be
inserted in the genome by a number of methods including
20 homologous recombination using a transfer vector, pTKgpt-OFIS
as described in Kaslow, et al., *Science* 252:1310-1313 (1991),
which is incorporated herein by reference.

Alternately the DNA encoding the SABP and DABP
binding domains may be inserted into another plasmid designed
25 for producing recombinant vaccinia, such as pGS62, Langford,
C.L., et al., 1986, *Mol. Cell. Biol.* 6:3191-3199. This
plasmid consists of a cloning site for insertion of foreign
genes, the P7.5 promoter of vaccinia to direct synthesis of
the inserted gene, and the vaccinia TK gene flanking both ends
30 of the foreign gene.

Confirmation of production of recombinant virus can
be achieved by DNA hybridization using CDNA encoding the DABP
and SABP binding domain polypeptides and by immunodetection
techniques using antibodies specific for the expressed binding
35 domain polypeptides. Virus stocks may be prepared by
infection of cells such as HELA S3 spinner cells and
harvesting of virus progeny.

The recombinant virus of the present invention can be used to induce anti-SABP and anti-DABP binding domain antibodies in mammals, such as mice or humans. In addition, the recombinant virus can be used to produce the SABP and DABP binding domains by infecting host cells *in vitro*, which in turn express the polypeptide (see section on expression of SABP and DABP binding domains in eukaryotic cells, above).

The present invention also relates to host cells infected with the recombinant virus. The host cells of the present invention are preferably mammalian, such as BSC-1 cells. Host cells infected with the recombinant virus express the DABP and SABP binding domains on their cell surfaces. In addition, membrane extracts of the infected cells induce protective antibodies when used to inoculate or boost previously inoculated mammals.

D. Purification of the SABP, DABP and EBL Binding Domain Polypeptides

The binding domain polypeptides produced by recombinant DNA technology may be purified by standard techniques well known to those of skill in the art. Recombinantly produced binding domain polypeptides can be directly expressed or expressed as a fusion protein. The protein is then purified by a combination of cell lysis (e. g., sonication) and affinity chromatography. For fusion products, subsequent digestion of the fusion protein with an appropriate proteolytic enzyme release the desired SABP and DABP binding domains.

The polypeptides of this invention may be purified to substantial purity by standard techniques well known in the art, including selective precipitation with such substances as ammonium sulfate, column chromatography, immunopurification methods, and others. See, for instance, R. Scopes, *Protein Purification: Principles and Practice*, Springer-Verlag: New York (1982), incorporated herein by reference.

E. Production of Binding Domains by protein chemistry techniques

The polypeptides of the invention can be synthetically prepared in a wide variety of ways. For

instance polypeptides of relatively short size, can be synthesized in solution or on a solid support in accordance with conventional techniques. Various automatic synthesizers are commercially available and can be used in accordance with known protocols. See, for example, Stewart and Young, *Solid Phase Peptide Synthesis*, 2d. ed., Pierce Chemical Co. (1984).

Alternatively, purified and isolated SABP, DABP or EBL family proteins may be treated with proteolytic enzymes in order to produce the binding domain polypeptides. For example, recombinant DABP and SABP proteins may be used for this purpose. The DABP and SABP protein sequence may then be analyzed to select proteolytic enzymes to be used to generate polypeptides containing desired regions of the DABP and SABP binding domain. The desired polypeptides are then purified by using standard techniques for protein and peptide purification. For a review of standard techniques see, *Methods in Enzymology*, "Guide to Protein Purification", M. Deutscher, ed. Vol. 182 (1990), pages 619-626, which is incorporated herein by reference.

F. Modification of nucleic acid and polypeptide sequences

The nucleotide sequences used to transfect the host cells used for production of recombinant binding domain polypeptides can be modified according to standard techniques to yield binding domain polypeptides, with a variety of desired properties. The binding domain polypeptides of the present invention can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the binding domain polypeptides can vary from the naturally-occurring sequence at the primary structure level by amino acid insertions, substitutions, deletions, and the like. These modifications can be used in a number of combinations to produce the final modified protein chain.

The amino acid sequence variants can be prepared with various objectives in mind, including facilitating purification and preparation of the recombinant polypeptides. The modified polypeptides are also useful for modifying plasma

half-life, improving therapeutic efficacy, and lessening the severity or occurrence of side effects during therapeutic use. The amino acid sequence variants are usually predetermined variants not found in nature but exhibit the same immunogenic activity as naturally occurring polypeptides. For instance, polypeptide fragments comprising only a portion (usually at least about 60-80%, typically 90-95%) of the primary structure may be produced. For use as vaccines, polypeptide fragments are typically preferred so long as at least one epitope capable of eliciting production of blocking antibodies remains.

In general, modifications of the sequences encoding the binding domain polypeptides may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Gilman and Smith, *Gene* 8:81-97 (1979) and Roberts, S. et al., *Nature* 328:731-734 (1987)). One of ordinary skill will appreciate that the effect of many mutations is difficult to predict. Thus, most modifications are evaluated by routine screening in a suitable assay for the desired characteristic. For instance, changes in the immunological character of the polypeptide can be detected by an appropriate competitive binding assay. Modifications of other properties such as redox or thermal stability, hydrophobicity, susceptibility to proteolysis, or the tendency to aggregate are all assayed according to standard techniques.

G. Diagnostic and Screening Assays

The polypeptides of the invention can be used in diagnostic applications for the detection of merozoites in a biological sample. The presence of parasites can be detected using several well recognized specific binding assays based on immunological results. (See U.S. Patents 4,366,241; 4,376,110; 4,517,288; and 4,837,168, which are hereby incorporated by reference.) For instance, labeled monoclonal antibodies to polypeptides of the invention can be used to detect merozoites in a biological sample. Alternatively, labelled polypeptides of the invention can be used to detect the presence of antibodies to SABP or DABP in a biological sample. For a review of the general procedures in diagnostic immunoassays,

see also *Basic and Clinical Immunology* 7th Edition (D. Stites and A. Terr ed.) 1991, which is hereby incorporated by reference.

In addition, modified polypeptides, antibodies or other compounds capable of inhibiting the interaction between SABP or DABP and erythrocytes can be assayed for biological activity. For instance, polypeptides can be recombinantly expressed on the surface of cells and the ability of the cells to bind erythrocytes can be measured as described below. Alternatively, peptides or antibodies can be tested for the ability to inhibit binding between erythrocytes and merozoites or SABP and DABP.

Cell-free assays can also be used to measure binding of DABP or SABP polypeptides to isolated Duffy antigen or glycophorin polypeptides. For instance, the erythrocyte proteins can be immobilized on a solid surface and binding of labelled SABP or DABP polypeptides can be measured.

Many assay formats employ labelled assay components. The labelling systems can be in a variety of forms. The label may be coupled directly or indirectly to the desired component of the assay according to methods well known in the art. A wide variety of labels may be used. The component may be labelled by any one of several methods. The most common method of detection is the use of autoradiography with ^3H , ^{125}I , ^{35}S , ^{14}C , or ^{32}P labelled compounds or the like.

Non-radioactive labels include ligands which bind to labelled antibodies, fluorophores, chemiluminescent agents, enzymes, and antibodies which can serve as specific binding pair members for a labelled ligand. The choice of label depends on sensitivity required, ease of conjugation with the compound, stability requirements, and available instrumentation.

In addition, the polypeptides of the invention can be assayed using animal models, well known to those of skill in the art. For *P. falciparum* the *in vivo* models include *Aotus* sp. monkeys or chimpanzees; for *P. vivax* the *in vivo* models include *Saimiri* monkeys.

H. Pharmaceutical compositions comprising binding domain polypeptides

The polypeptides of the invention are useful in therapeutic and prophylactic applications for the treatment of malaria. Pharmaceutical compositions of the invention are suitable for use in a variety of drug delivery systems.

5 Suitable formulations for use in the present invention are found in *Remington's Pharmaceutical Sciences*, Mack Publishing Company, Philadelphia, PA, 17th ed. (1985), which is incorporated herein by reference. For a brief review of methods for drug delivery, see, Langer, *Science* 249:1 527-1533
10 (1990), which is incorporated herein by reference.

The polypeptides of the present invention can be used in pharmaceutical and vaccine compositions that are useful for administration to mammals, particularly humans. The polypeptides can be administered together in certain
15 circumstances, e.g. where infection by both *P. falciparum* and *P. vivax* is likely. Thus, a single pharmaceutical composition can be used for the treatment or prophylaxis of malaria caused by both parasites.

The compositions are suitable for single
20 administrations or a series of administrations. When given as a series, inoculations subsequent to the initial administration are given to boost the immune response and are typically referred to as booster inoculations.

The pharmaceutical compositions of the invention are
25 intended for parenteral, topical, oral or local administration. Preferably, the pharmaceutical compositions are administered parenterally, e.g., intravenously, subcutaneously, intradermally, or intramuscularly. Thus, the invention provides compositions for parenteral administration
30 that comprise a solution of the agents described above dissolved or suspended in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers may be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be
35 sterilized by conventional, well known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile solution

prior to administration. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions, such as pH adjusting and buffering agents, tonicity adjusting agents, wetting agents and the like, for example, sodium acetate, sodium lactate, sodium chloride, potassium chloride, calcium chloride, sorbitan monolaurate, triethanolamine oleate, etc.

For solid compositions, conventional nontoxic solid carriers may be used which include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talcum, cellulose, glucose, sucrose, magnesium carbonate, and the like. For oral administration, a pharmaceutically acceptable nontoxic composition is formed by incorporating any of the normally employed excipients, such as those carriers previously listed, and generally 10-95% of active ingredient and more preferably at a concentration of 25%-75%.

For aerosol administration, the polypeptides are preferably supplied in finely divided form along with a surfactant and propellant. The surfactant must, of course, be nontoxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed. A carrier can also be included, as desired, as with, e.g., lecithin for intranasal delivery.

In certain embodiments patients with malaria may be treated with SABP or DABP polypeptides or other specific blocking agents (e.g. monoclonal antibodies) that prevent binding of *Plasmodium* merozoites and schizonts to the erythrocyte surface.

The amount administered to the patient will vary depending upon what is being administered, the state of the patient and the manner of administration. In therapeutic applications, compositions are administered to a patient

already suffering from malaria in an amount sufficient to inhibit spread of the parasite through erythrocytes and thus cure or at least partially arrest the symptoms of the disease and its complications. An amount adequate to accomplish this is defined as "therapeutically effective dose." Amounts effective for this use will depend on the severity of the disease and the weight and general state of the patient. But will generally be in the range of about 1mg to about 5gm per day, preferably about 100 mg per day, for a 70 kg patient.

Alternatively, the polypeptides of the invention can be used prophylactically as vaccines. The vaccines of the invention contain as an active ingredient an immunogenically effective amount of the binding domain polypeptide or of a recombinant virus as described herein. The immune response may include the generation of antibodies; activation of cytotoxic T lymphocytes (CTL) against cells presenting peptides derived from the peptides encoded by the SABP, DABP or EBL sequences of the present invention, or other mechanisms well known in the art. See e.g. Paul *Fundamental Immunology Second Edition* published by Raven press New York (incorporated herein by reference) for a description of immune response. Useful carriers are well known in the art, and include, for example, thyroglobulin, albumins such as human serum albumin, tetanus toxoid, polyamino acids such as poly(D-lysine:D-glutamic acid), influenza, hepatitis B virus core protein, hepatitis B virus recombinant vaccine. The vaccines can also contain a physiologically tolerable (acceptable) diluent such as water, phosphate buffered saline, or saline, and further typically include an adjuvant. Adjuvants such as incomplete Freund's adjuvant, aluminum phosphate, aluminum hydroxide, or alum are materials well known in the art.

The DNA or RNA encoding the SABP or DABP binding domains and the EBL gene family motifs may be introduced into patients to obtain an immune response to the polypeptides which the nucleic acid encodes. Wolff et. al., *Science* 247: 1465-1468 (1990) which is incorporated herein by reference

describes the use of nucleic acids to produce expression of the genes which the nucleic acids encode.

Vaccine compositions containing the polypeptides, nucleic acids or viruses of the invention are administered to a patient to elicit a protective immune response against the polypeptide. A "protective immune response" is one which prevents or inhibits the spread of the parasite through erythrocytes and thus at least partially prevent the symptoms of the disease and its complications. An amount sufficient to accomplish this is defined as an "immunogenically effective dose." Amounts effective for this use will depend on the composition, the manner of administration, the weight and general state of health of the patient, and the judgment of the prescribing physician. For peptide compositions, the general range for the initial immunization (that is for therapeutic or prophylactic administration) is from about 100 μ g to about 1 gm of peptide for a 70 kg patient, followed by boosting dosages of from about 100 μ g to about 1 gm of the polypeptide pursuant to a boosting regimen over weeks to months depending upon the patient's response and condition e.g. by measuring levels of parasite in the patient's blood. For nucleic acids, typically 30-1000ug of nucleic acid is injected into a 70kg patient, more typically about 50-150ug of nucleic acid is injected into a 70kg patient followed by boosting doses as appropriate.

The following example is offered by way of illustration, not by way of limitation.

EXAMPLE

Identification of the amino-terminal, cysteine-rich region of SABP and DABP as binding domains for erythrocytes

5

1. Expression of the SABP binding domain polypeptide on the surface of Cos cells.

To demonstrate that the amino-terminal, cysteine-rich region of the SABP protein is the sialic acid binding region, this region of the protein was expressed on the surface of mammalian Cos cells *in vitro*. This DNA sequence is from position 1 to position 1848 of the SABP DNA sequence (SEQ ID No 3). Polymerase chain reaction technology (PCR) was used to amplify this region of the SABP DNA directly from the cloned gene.

Sequences corresponding to restriction endonuclease sites for PvuII or ApaI were incorporated into the oligonucleotide sequence of the probes used in PCR amplification in order to facilitate insertion of the PCR-amplified regions into the pRE4 vector (see below). The specific oligonucleotides, 5'-ATCGATCAGCTGGGAAGAAATACTTCATCT-3' and 5'-ATCGATGGGCCCCGAAGTTTGTTCATTATT-3' were synthesized. These oligonucleotides were used as primers to PCR-amplify the region of the DNA sequence encoding the cysteine-rich amino terminal region of the SABP protein.

PCR conditions were based on the standard described in Saiki, et al., Science 239: 487-491 (1988). Template DNA was provided from cloned fragments of the gene encoding SABP which had been spliced and re-cloned as a single open-reading frame piece.

The vector, pRE4, used for expression in Cos cells is shown in Fig.1. The vector has an SV40 origin of replication, an ampicillin resistance marker and the Herpes simplex virus glycoprotein D gene (HSV glyD) cloned downstream of the Rous sarcoma virus long terminal repeats (RSV LTR). Part of the extracellular domain of the HSV glyD gene was excised using the PvuII and ApaI sites in HSV glyD.

As described above, the PCR oligonucleotide primers contained the PvuII or ApaI restriction sites. The PCR-amplified DNA fragments obtained above were digested with the restriction enzymes PvuII and ApaI and cloned into the PvuII and ApaI sites of the vector pRE4. These constructs were designed to express regions of the SABP protein as chimeric proteins with the signal sequence of HSV glyD at the N-terminal end and the transmembrane and cytoplasmic domain of HSV glyD at the C-terminal end. The signal sequence of HSV glyD targets these chimeric proteins to the surface of Cos cells and the transmembrane segment of HSV glyD anchors these chimeric proteins to the Cos cell surface.

Mammalian Cos cells were transfected with the pRE4 constructs containing the PCR-amplified SABP DNA regions, by calcium phosphate precipitation according to standard techniques.

2. Expression of the DABP binding domain polypeptide on the surface of Cos cells.

To demonstrate that the amino-terminal, cysteine-rich region of the DABP protein is the binding domain, this region was expressed on the surface of Cos cells. This region of the DNA sequence from position 1-975 was first PCR-amplified (SEQ ID No 1).

Sequences corresponding to restriction endonuclease sites for PvuII or ApaI were incorporated into the oligonucleotide probes used for PCR amplification in order to facilitate subsequent insertion of the amplified DNA into the pRE4 vector, as described above. The oligonucleotides, 5'-TCTCGTCAGCTGACGATCTCTAGTGCTATT-3' and 5'-ACGAGTGGGCCCTGTCACAACTTCCTGAGT-3' were synthesized. These oligonucleotides were used as primers to amplify the region of the DABP DNA sequence encoding the cysteine-rich, amino-terminal region of the DABP protein directly from the cloned DABP gene, using the same conditions described above.

The same pRE4 vector described above in the section on expression of SABP regions in Cos cells was also used as a vector for the DABP DNA regions.

3. Binding studies with erythrocytes.

To demonstrate their ability to bind human erythrocytes, the transfected Cos cells expressing binding domains from DABP and SABP were incubated with erythrocytes for two hours at 37°C in culture media (DMEM/10% FBS). The non-adherent erythrocytes were removed with five washes of phosphate-buffered saline and the bound erythrocytes were observed by light microscopy. Cos cells expressing the amino terminal, cysteine-rich SABP polypeptides on their surface bound untreated human erythrocytes, but did not bind neuraminidase treated erythrocytes, that is, erythrocytes which lack sialic acid residues on their surface (data not shown). Cos cells expressing other regions of the SABP protein on their surface did not bind human erythrocytes (data not shown). These results identified the amino-terminal, cysteine-rich region of SABP as the erythrocyte binding domain and indicated that the binding of Cos cells expressing these regions to human erythrocytes is specific. Furthermore, the binding of the expressed region to erythrocytes is identical to the binding pattern seen for the authentic SABP- 175 molecule upon binding to erythrocytes.

Similarly, Cos cells expressing the amino-terminal cysteine-rich region of DABP on their surface bound Duffy-positive human erythrocytes, but did not bind Duffy-negative human erythrocytes, that is erythrocytes which lack the Duffy blood group antigen (data not shown). Cos cells expressing other regions of the DABP protein on their surface did not bind human erythrocytes (data not shown). These results identified the amino-terminal cysteine rich region of DABP as the erythrocyte binding domain and indicated that the binding of the Cos cells was specific.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT:
 - (A) NAME: The United States Of America,
as represented by
The Secretary of the Department
of Health and Human Services
- (ii) TITLE OF INVENTION: BINDING DOMAINS FROM PLASMODIUM VIVAX
AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING PROTEINS
- (iii) NUMBER OF SEQUENCES: 12
- (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (v) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: WO not yet assigned
 - (B) FILING DATE: 07-SEP-1994
 - (C) CLASSIFICATION:
- (vi) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/119,677
 - (B) FILING DATE: 10-SEP-1993
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Bastian, Kevin L.
 - (B) REGISTRATION NUMBER: 34,774
 - (C) REFERENCE/DOCKET NUMBER: 15280-139000
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (415) 543-9600
 - (B) TELEFAX: (415) 543-5043

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4084 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Plasmodium vivax
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

AAGCTTTTAA AAATAGCAAC AAAATTTCTGA AACATTGCCA CAAAATTTT ATGTTTTACA	60
TATATTTAGA TTCATACAAT TTAGGTGTAC CCTGTTTTTT GATATATGCG CTAAATTTT	120
TTTTTCGCTC ATATGTTTAG TTATATGTGT AGAACAACTT GCTGAATAAA TTACGTACAC	180

TTTCTGTTCT	GAATAATATT	ACCACATACA	TTTAATTTTA	AATACTATGA	AAGGAAAAAA	240
CCGCTCTTTA	TTTGTTCCTC	TAGTTTTATT	ATTGTTACAC	AAGGTATCAT	ATAAGGATGA	300
TTTTTCTATC	ACACTAATAA	ATTATCATGA	AGGAAAAAAA	TATTTAATTA	TACTAAAAAG	360
AAAATTAGAA	AAAGCTAATA	ATCGTGATGT	TGCAATTTT	TTTCTTCATT	TCTCTCAGGT	420
AAATAATGTA	TTATTAGAAC	GAACAATTGA	AACCCCTCTA	GAATGCAAAA	ATGAATATGT	480
GAAAGGTGAA	AATGGTTATA	AATTAGCTAA	AGGACACCAC	TGTGTTGAGG	AAGATAACTT	540
AGAACGATGG	TTACAAGGAA	CCAATGAAAG	AAGAAGTGAG	GAAAATATAA	AATATAAATA	600
TGGAGTAACG	GAATAAAAAA	TAAAGTATGC	GCAATGAAT	GGAAAAAGAA	GCAGCCGCAT	660
TTTGAAGGAA	TCAATTTACG	GGGCGCATAA	CTTTGGAGGC	AACAGTTACA	TGGAGGGAAA	720
AGATGGAGGA	GATAAAACTG	GGGAGGAAAA	AGATGGAGAA	CATAAACTG	ATAGTAAAC	780
TGATAACGGG	AAAGGTGCAA	ACAATTTGGT	AATGTTAGAT	TATGAGACAT	CTAGCAATGG	840
CCAGCCAGCG	GGAACCCCTG	ATAATGTTCT	TGAATTTGTG	ACTGGGCATG	AGGGAAATTC	900
TCGTAAAAAT	TCCTCGAATG	GTGGCAATCC	TTACGATATT	GATCATAAGA	AAACGATCTC	960
TAGTGCTATT	ATAAATCATG	CTTTTCTTCA	AAATACTGTA	ATGAAAAACT	GTAATTATAA	1020
GAGAAAACGT	CGGGAAAGAG	ATTGGGACTG	TAACACTAAG	AAGGATGTTT	GTATACCAGA	1080
TCGAAGATAT	CAATTATGTA	TGAAGGAACT	TACGAATTTG	GTAAATAATA	CAGACACAAA	1140
TTTTCATAGG	GATATAACAT	TTCGAAAATT	ATATTTGAAA	AGGAAACTTA	TTTATGATGC	1200
TGCAGTAGAG	GGCGATTTAT	TACTTAAGTT	GAATAACTAC	AGATATAACA	AAGACTTTTG	1260
CAAGGATATA	AGATGGAGTT	TGGGAGATTT	TGGAGATATA	ATTATGGGAA	CGGATATGGA	1320
AGGCATCGGA	TATTCCAAAG	TAGTGGA AAA	TAATTTGCCG	AGCATCTTTG	GAACTGATGA	1380
AAAGGCCCAA	CAGCGTCGTA	AACAGTG GTG	GAATGAATCT	AAAGCACAAA	TTTGGACAGC	1440
AATGATGTAC	TCAGTTAAAA	AAAGATTAAA	GGGGAATTTT	ATATGGATTT	GTAAATTAAA	1500
TGTTGCGGTA	AATATAGAAC	CGCAGATATA	TAGATGGATT	CGAGAATGGG	GAAGGGATTA	1560
CGTGT CAGAA	TTGCCACAG	AAGTGCAAAA	ACTGAAAGAA	AAATGTGATG	GAAAAATCAA	1620
TTATACTGAT	AAAAAAGTAT	GTAAGGTACC	ACCATGTCAA	AATGCGTGTA	AATCATATGA	1680
TCAATGGATA	ACCAGAAAAA	AAAATCAATG	GGATGTTCTG	TCAAATAAAT	TCATAAGTGT	1740
AAAAAACGCA	GAAAAGGTTT	AGACGGCAGG	TATCGTAACT	CCTTATGATA	TACTAAAACA	1800
GGAGTTAGAT	GAATTTAACG	AGGTGGCTTT	TGAGAATGAA	ATTAACAAAC	GTGATGGTGC	1860
ATATATTGAG	TTATGCGTTT	GTTCCGTTGA	AGAGGCTAAA	AAAAATACTC	AGGAAGTTGT	1920
GACAAATGTG	GACAATGCTG	CTAAATCTCA	GGCCACCAAT	TCAAATCCGA	TAAGTCAGCC	1980
TGTAGATAGT	AGTAAAGCGG	AGAAGGTTCC	AGGAGATTCT	ACGCATGGAA	ATGTTAACAG	2040
TGGCCAAGAT	AGTTCTACCA	CAGGTAAAGC	TGTTACGGGG	GATGGTCAAA	ATGGAAATCA	2100
GACACCTGCA	GAAAGCGATG	TACAGCGAAG	TGATATTGCC	GAAAGTGTA	GTGCTAAAAA	2160
TGTTGATCCG	CAGAAATCTG	TAAGTAAAAG	AAGTGACGAC	ACTGCAAGCG	TTACAGGTAT	2220

TGCCGAAGCT	GGAAAGGAAA	ACTTAGGCGC	ATCAAATAGT	CGACCTTCTG	AGTCCACCGT	2280
TGAAGCAAAT	AGCCCAGGTG	ATGATACTGT	GAACAGTGCA	TCTATACCTG	TAGTGAGTGG	2340
TGAAAACCCA	TTGGTAACCC	CCTATAATGG	TTTGAGGCAT	TCGAAAGACA	ATAGTGATAG	2400
CGATGGACCT	GCGGAATCAA	TGGCGAATCC	TGATTCAAAT	AGTAAAGGTG	AGACGGGAAA	2460
GGGGCAAGAT	AATGATATGG	CGAAGGCTAC	TAAAGATAGT	AGTAATAGTT	CAGATGGTAC	2520
CAGCTCTGCT	ACGGGTGATA	CTACTGATGC	AGTTGATAGG	GAAATTAATA	AAGGTGTTCC	2580
TGAGGATAGG	GATAAAACTG	TAGGAAGTAA	AGATGGAGGG	GGGGAAGATA	ACTCTGCAAA	2640
TAAGGATGCA	GCGACTGTAG	TTGGTGAGGA	TAGAATTCGT	GAGAACAGCG	CTGGTGGTAG	2700
CACTAATGAT	AGATCAAAAA	ATGACACGGA	AAAGAACGGG	GCCTCTACCC	CTGACAGTAA	2760
ACAAAGTGAG	GATGCAACTG	CGCTAAGTAA	AACCGAAAGT	TTAGAATCAA	CAGAAAGTGG	2820
AGATAGAACT	ACTAATGATA	CAACTAACAG	TTTAGAAAAT	AAAAATGGAG	GAAAAGAAAA	2880
GGATTTACAA	AAGCATGATT	TTAAAAGTAA	TGATACGCCG	AATGAAGAAC	CAAATTCTGA	2940
TCAAACACTA	GATGCAGAAG	GACATGACAG	GGATAGCATC	AAAAATGATA	AAGCAGAAAG	3000
GAGAAAGCAT	ATGAATAAAG	ATACTTTTAC	GAAAAATACA	AATAGTCACC	ATTTAAATAG	3060
TAATAATAAT	TTGAGTAATG	GAAAATTAGA	TATAAAAGAA	TACAAATACA	GAGATGTCAA	3120
AGCAACAAGG	GAAGATATTA	TATTAATGTC	TTCAGTACGC	AAGTGCAACA	ATAATATTTT	3180
TTTAGAGTAC	TGTAACCTCTG	TAGAGGACAA	AATATCATCG	AATACTTGTT	CTAGAGAGAA	3240
AAGTAAAAAT	TTATGTTGCT	CAATATCGGA	TTTTTGTTTG	AACTATTTTG	ACGTGTATTC	3300
TTATGAGTAT	CTTAGCTGCA	TGAAAAAGGA	ATTGGAAGAT	CCATCCTACA	AGTGCTTTAC	3360
GAAAGGGGGC	TTTAAAGGTA	TGCAGAAAAA	GATGCTGAAT	AGAGAAAGGT	GTTGAGTAAA	3420
TTAAAAAGGA	ATTAATTTTA	GGAATGTTAT	AAACATTTTT	GTACCCAAAA	TTCTTTTTGC	3480
AGACAAGACT	TACTTTGCCG	CGGCGGGAGC	GTTGCTGATA	CTGCTGTTGT	TAATTGCTTC	3540
AAGGAAGATG	ATCAAAAATG	AGTAACCAGA	AAATAAAATA	AAATAACATA	AAATAAAATA	3600
AAAAC TAGAA	TAACAATTAA	AATAAAATAA	AATGAGAAAT	GCCTGTTAAT	GCACAGTTAA	3660
TTCTAACGAT	TCCATTTGTG	AAGTTTTTAA	GAGAGCACAA	ATGCATAGTC	ATTATGTCCA	3720
TGCATATATA	CACATATATG	TACGTATATA	TAATAAACGC	ACACTTTCTT	GTTTCGTACAG	3780
TTCTGAAGAA	GCTACATTTA	ATGAGTTTGA	AGAATACTGT	GATAATATTC	ACAGAATCCC	3840
TCTGATGCCT	AACAGTAATT	CAAATTTCAA	GAGCAAAATT	CCATTTAAAA	AGAAATGTTA	3900
CATCATTTTG	CGTTTTTCTT	TTTTTCTTTT	TTTTTCTTTT	TTTAGATATT	GAACACATGC	3960
AGCCATCAAC	CCCCCTGGAT	TATTCATGAT	GCTACTTTGG	TAAGTAAAAG	CAATTCTGAT	4020
TGTAGTGCTG	ATGTAATTTT	AGTCATTTTG	CTTGCTGCAA	TAAACGAGAA	AATATATCAA	4080
GCTT						4084

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1115 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Plasmodium vivax

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Met Lys Gly Lys Asn Arg Ser Leu Phe Val Leu Leu Val Leu Leu Leu
1           5           10           15
Leu His Lys Val Ser Tyr Lys Asp Asp Phe Ser Ile Thr Leu Ile Asn
20           25           30
Tyr His Glu Gly Lys Lys Tyr Leu Ile Ile Leu Lys Arg Lys Leu Glu
35           40           45
Lys Ala Asn Asn Arg Asp Val Cys Asn Phe Phe Leu His Phe Ser Gln
50           55           60
Val Asn Asn Val Leu Leu Glu Arg Thr Ile Glu Thr Leu Leu Glu Cys
65           70           75           80
Lys Asn Glu Tyr Val Lys Gly Glu Asn Gly Tyr Lys Leu Ala Lys Gly
85           90           95
His His Cys Val Glu Glu Asp Asn Leu Glu Arg Trp Leu Gln Gly Thr
100          105          110
Asn Glu Arg Arg Ser Glu Glu Asn Ile Lys Tyr Lys Tyr Gly Val Thr
115          120          125
Glu Leu Lys Ile Lys Tyr Ala Gln Met Asn Gly Lys Arg Ser Ser Arg
130          135          140
Ile Leu Lys Glu Ser Ile Tyr Gly Ala His Asn Phe Gly Gly Asn Ser
145          150          155          160
Tyr Met Glu Gly Lys Asp Gly Gly Asp Lys Thr Gly Glu Glu Lys Asp
165          170          175
Gly Glu His Lys Thr Asp Ser Lys Thr Asp Asn Gly Lys Gly Ala Asn
180          185          190
Asn Leu Val Met Leu Asp Tyr Glu Thr Ser Ser Asn Gly Gln Pro Ala
195          200          205
Gly Thr Leu Asp Asn Val Leu Glu Phe Val Thr Gly His Glu Gly Asn
210          215          220
Ser Arg Lys Asn Ser Ser Asn Gly Gly Asn Pro Tyr Asp Ile Asp His
225          230          235          240
Lys Lys Thr Ile Ser Ser Ala Ile Il Asn His Ala Phe Leu Gln Asn
245          250          255

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Thr Val Met Lys Asn Cys Asn Tyr Lys Arg Lys Arg Arg Glu Arg Asp
 260 265 270
 Trp Asp Cys Asn Thr Lys Lys Asp Val Cys Ile Pro Asp Arg Arg Tyr
 275 280 285
 Gln Leu Cys Met Lys Glu Leu Thr Asn Leu Val Asn Asn Thr Asp Thr
 290 295 300
 Asn Phe His Arg Asp Ile Thr Phe Arg Lys Leu Tyr Leu Lys Arg Lys
 305 310 315 320
 Leu Ile Tyr Asp Ala Ala Val Glu Gly Asp Leu Leu Leu Lys Leu Asn
 325 330 335
 Asn Tyr Arg Tyr Asn Lys Asp Phe Cys Lys Asp Ile Arg Trp Ser Leu
 340 345 350
 Gly Asp Phe Gly Asp Ile Ile Met Gly Thr Asp Met Glu Gly Ile Gly
 355 360 365
 Tyr Ser Lys Val Val Glu Asn Asn Leu Arg Ser Ile Phe Gly Thr Asp
 370 375 380
 Glu Lys Ala Gln Gln Arg Arg Lys Gln Trp Trp Asn Glu Ser Lys Ala
 385 390 395 400
 Gln Ile Trp Thr Ala Met Met Tyr Ser Val Lys Lys Arg Leu Lys Gly
 405 410 415
 Asn Phe Ile Trp Ile Cys Lys Leu Asn Val Ala Val Asn Ile Glu Pro
 420 425 430
 Gln Ile Tyr Arg Trp Ile Arg Glu Trp Gly Arg Asp Tyr Val Ser Glu
 435 440 445
 Leu Pro Thr Glu Val Gln Lys Leu Lys Glu Lys Cys Asp Gly Lys Ile
 450 455 460
 Asn Tyr Thr Asp Lys Lys Val Cys Lys Val Pro Pro Cys Gln Asn Ala
 465 470 475 480
 Cys Lys Ser Tyr Asp Gln Trp Ile Thr Arg Lys Lys Asn Gln Trp Asp
 485 490 495
 Val Leu Ser Asn Lys Phe Ile Ser Val Lys Asn Ala Glu Lys Val Gln
 500 505 510
 Thr Ala Gly Ile Val Thr Pro Tyr Asp Ile Leu Lys Gln Glu Leu Asp
 515 520 525
 Glu Phe Asn Glu Val Ala Phe Glu Asn Glu Ile Asn Lys Arg Asp Gly
 530 535 540
 Ala Tyr Ile Glu Leu Cys Val Cys Ser Val Glu Glu Ala Lys Lys Asn
 545 550 555 560
 Thr Gln Glu Val Val Thr Asn Val Asp Asn Ala Ala Lys Ser Gln Ala
 565 570 575
 Thr Asn Ser Asn Pro Ile Ser Gln Pro Val Asp Ser Ser Lys Ala Glu
 580 585 590
 Lys Val Pro Gly Asp Ser Thr His Gly Asn Val Asn Ser Gly Gln Asp
 595 600 605

Ser Ser Thr Thr Gly Lys Ala Val Thr Gly Asp Gly Gln Asn Gly Asn
 610 615 620
 Gln Thr Pro Ala Glu Ser Asp Val Gln Arg Ser Asp Il Ala Glu Ser
 625 630 635 640
 Val Ser Ala Lys Asn Val Asp Pro Gln Lys Ser Val Ser Lys Arg Ser
 645 650 655
 Asp Asp Thr Ala Ser Val Thr Gly Ile Ala Glu Ala Gly Lys Glu Asn
 660 665 670
 Leu Gly Ala Ser Asn Ser Arg Pro Ser Glu Ser Thr Val Glu Ala Asn
 675 680 685
 Ser Pro Gly Asp Asp Thr Val Asn Ser Ala Ser Ile Pro Val Val Ser
 690 695 700
 Gly Glu Asn Pro Leu Val Thr Pro Tyr Asn Gly Leu Arg His Ser Lys
 705 710 715 720
 Asp Asn Ser Asp Ser Asp Gly Pro Ala Glu Ser Met Ala Asn Pro Asp
 725 730 735
 Ser Asn Ser Lys Gly Glu Thr Gly Lys Gly Gln Asp Asn Asp Met Ala
 740 745 750
 Lys Ala Thr Lys Asp Ser Ser Asn Ser Ser Asp Gly Thr Ser Ser Ala
 755 760 765
 Thr Gly Asp Thr Thr Asp Ala Val Asp Arg Glu Ile Asn Lys Gly Val
 770 775 780
 Pro Glu Asp Arg Asp Lys Thr Val Gly Ser Lys Asp Gly Gly Gly Glu
 785 790 795 800
 Asp Asn Ser Ala Asn Lys Asp Ala Ala Thr Val Val Gly Glu Asp Arg
 805 810 815
 Ile Arg Glu Asn Ser Ala Gly Gly Ser Thr Asn Asp Arg Ser Lys Asn
 820 825 830
 Asp Thr Glu Lys Asn Gly Ala Ser Thr Pro Asp Ser Lys Gln Ser Glu
 835 840 845
 Asp Ala Thr Ala Leu Ser Lys Thr Glu Ser Leu Glu Ser Thr Glu Ser
 850 855 860
 Gly Asp Arg Thr Thr Asn Asp Thr Thr Asn Ser Leu Glu Asn Lys Asn
 865 870 875 880
 Gly Gly Lys Glu Lys Asp Leu Gln Lys His Asp Phe Lys Ser Asn Asp
 885 890 895
 Thr Pro Asn Glu Glu Pro Asn Ser Asp Gln Thr Thr Asp Ala Glu Gly
 900 905 910
 His Asp Arg Asp Ser Ile Lys Asn Asp Lys Ala Glu Arg Arg Lys His
 915 920 925
 Met Asn Lys Asp Thr Phe Thr Lys Asn Thr Asn Ser His His Leu Asn
 930 935 940
 Ser Asn Asn Asn Leu Ser Asn Gly Lys Leu Asp Ile Lys Glu Tyr Lys
 945 950 955 960

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Tyr Arg Asp Val Lys Ala Thr Arg Glu Asp Ile Ile Leu Met Ser Ser
 965 970 975
 Val Arg Lys Cys Asn Asn Asn Ile Ser Leu Glu Tyr Cys Asn Ser Val
 980 985 990
 Glu Asp Lys Ile Ser Ser Asn Thr Cys Ser Arg Glu Lys Ser Lys Asn
 995 1000 1005
 Leu Cys Cys Ser Ile Ser Asp Phe Cys Leu Asn Tyr Phe Asp Val Tyr
 1010 1015 1020
 Ser Tyr Glu Tyr Leu Ser Cys Met Lys Lys Glu Phe Glu Asp Pro Ser
 1025 1030 1035 1040
 Tyr Lys Cys Phe Thr Lys Gly Gly Phe Lys Ile Asp Lys Thr Tyr Phe
 1045 1050 1055
 Ala Ala Ala Gly Ala Leu Leu Ile Leu Leu Leu Ile Ala Ser Arg Lys
 1060 1065 1070
 Met Ile Lys Asn Asp Ser Glu Glu Ala Thr Phe Asn Glu Phe Glu Glu
 1075 1080 1085
 Tyr Cys Asp Asn Ile His Arg Ile Pro Leu Met Pro Asn Asn Ile Glu
 1090 1095 1100
 His Met Gln Pro Ser Thr Pro Leu Asp Tyr Ser
 1105 1110 1115

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4507 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

TATATATATA TATATATATA GATAATAACA TATAATATA TTCAATGTGC ATACAATGAA	60
ATGTAATATT AGTATATATT TTTTGTCTC CTCTTTGTG TTATATTTTG CAAAAGCTAG	120
GAATGAATAT GATATAAAAG AGAATGAAAA ATTTTGTAGC GTGTATAAAG AAAAATTTAA	180
TGAATTAGAT AAAAAGAAAT ATGGAAATGT TCAAAAACT GATAAGAAAA TATTTACTTT	240
TATAGAAAAT AAATTAGATA TTTTAAATAA TTCAAATTT AATAAAAGAT GGAAGAGTTA	300
TGGAACCTCA GATAATATAG ATAAAAATAT GTCTTTAATA AATAAACATA ATAATGAAGA	360
AATGTTTAAC AACAAATTATC AATCATTTTT ATCGACAAGT TCATTAATAA AGCAAAATAA	420
ATATGTTTCT ATTAACGCTG TACGTGTGTC TAGGATATTA AGTTTCCTGG ATTCTAGAAT	480
TAATAATGGA AGAAATACTT CATCTAATAA CGAAGTTTTA AGTAATTGTA GGGAAAAAAG	540
GAAAGGAATG AAATGGGATT GTAAAAAGAA AAATGATAGA AGCAACTATG TATGTATTCC	600

TGATCGTAGA	ATCCAATTAT	GCATTGTTAA	TCTTAGCATT	ATTAAAACAT	ATACAAAAGA	660
GACCATGAAG	GATCATTTCA	TTGAAGCCTC	TAAAAAAGAA	TCTCAACTTT	TGCTTAAAAA	720
AAATGATAAC	AAATATAATT	CTAAATTTTG	TAATGATTTG	AAGAATAGTT	TTTTAGATTA	780
TGGACATCTT	GCTATGGGAA	ATGATATGGA	TTTTGGAGGT	TATTCAACTA	AGGCAGAAAA	840
CAAATTCAA	GAAGTTTTTA	AAGGGGCTCA	TGGGGAAATA	AGTGAACATA	AAATTAAAAA	900
TTTTAGAAAA	GAATGGTGGA	ATGAATTTAG	AGAGAACTT	TGGGAAGCTA	TGTTATCTGA	960
GCATAAAAAT	AATATAAATA	ATTGTAAAAA	TATTCCCCAA	GAAGAATTAC	AAATTACTCA	1020
ATGGATAAAA	GAATGGCATG	GAGAATTTTT	GCTTGAAAGA	GATAATAGAT	CAAATTGCC	1080
AAAAAGTAAA	TGTAAAAATA	ATACATTATA	TGAAGCATGT	GAGAAGGAAT	GTATTGATCC	1140
ATGTATGAAA	TATAGAGATT	GGATTATTAG	AAGTAAATTT	GAATGGCATA	CGTTATCGAA	1200
AGAATATGAA	ACTCAAAAAG	TTCCAAAGGA	AAATGCGGAA	AATTATTTAA	TCAAAATTTT	1260
AGAAAACAAG	AATGATGCTA	AAGTAAGTTT	ATTATTGAAT	AATTGTGATG	CTGAATATTC	1320
AAAATATTGT	GATTGTAAAC	ATACTACTAC	TCTCGTTAAA	AGCGTTTTAA	ATGGTAACGA	1380
CAATACAATT	AAGGAAAAGC	GTGAACATAT	TGATTTAGAT	GATTTTCTA	AATTTGGATG	1440
TGATAAAAAT	TCCGTTGATA	CAAACACAAA	GGTGTGGGAA	TGTAAAAACC	CTTATATATT	1500
ATCCACTAAA	GATGTATGTG	TACCTCCGAG	GAGGCAAGAA	TTATGTCTTG	GAAACATTGA	1560
TAGAATATAC	GATAAAAACC	TATTAATGAT	AAAAGAGCAT	ATTCTTGCTA	TTGCAATATA	1620
TGAATCAAGA	ATATTGAAAC	GAAAATATAA	GAATAAGAT	GATAAAGAAG	TTTGTAATAT	1680
CATAAATAAA	ACTTTCGCTG	ATATAAGAGA	TATTATAGGA	GGTACTGATT	ATTGGAATGA	1740
TTTGAGCAAT	AGAAAATTAG	TAGGAAAAAT	TAACACAAAT	TCAAAATATG	TTCACAGGAA	1800
TAAAAAAAAT	GATAAGCTTT	TTCGTGATGA	GTGGTGGAAA	GTTATTAAAA	AAGATGTATG	1860
GAATGTGATA	TCATGGGTAT	TCAAGGATAA	AACTGTTTGT	AAAGAAGATG	ATATTGAAAA	1920
TATACCACAA	TTCTTCAGAT	GGTTTAGTGA	ATGGGGTGAT	GATTATTGCC	AGGATAAAAC	1980
AAAAATGATA	GAGACTCTGA	AGGTTGAATG	CAAAGAAAAA	CCTTGTGAAG	ATGACAATTG	2040
TAAAAGTAAA	TGTAATTCAT	ATAAGAATG	GATATCAAAA	AAAAAAGAAG	AGTATAATAA	2100
ACAAGCCAAA	CAATACCAAG	AATATCAAAA	AGGAAATAAT	TACAAAATGT	ATTCTGAATT	2160
TAAATCTATA	AAACCAGAAG	TTTATTTAAA	GAAATACTCG	GAAAAATGTT	CTAACCTAAA	2220
TTTCGAAGAT	GAATTTAAGG	AAGAATTACA	TTCAGATTAT	AAAAATAAAT	GTACGATGTG	2280
TCCAGAAGTA	AAGGATGTAC	CAATTTCTAT	AATAAGAAAT	AATGAACAAA	CTTCGCAAGA	2340
AGCAGTTCCT	GAGGAAAACA	CTGAAATAGC	ACACAGAACG	GAAACTCCAT	CTATCTCTGA	2400
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ATCACCAGAA	AATTCAAGAC	CTGAACTGA	TGCTAAAGAT	ACTTCTAACT	TGTTAAAAAT	2520
AAAAGGAGAT	GTTGATATTA	GTATGCCTAA	AGCAGTTATT	GGGAGCAGTC	CTAATGATAA	2580
TATAAATGTT	ACTGAACAAG	GGGATAATAT	TTCCGGGGTG	AATTCTAAAC	CTTTATCTGA	2640

TGATGTACGT	CCAGATAAAA	AGGAATTAGA	AGATCAAAAT	AGTGATGAAT	CGGAAGAAAC	2700
TGTAGTAAAT	CATATATCAA	AAAGTCCATC	TATAAATAAT	GGAGATGATT	CAGGCAGTGG	2760
AAGTGCAACA	GTGAGTGAAT	CTAGTAGTTC	AAATACTGGA	TTGTCTATTG	ATGATGATAG	2820
AAATGGTGAT	ACATTTGTTC	GAACACAAGA	TACAGCAAAT	ACTGAAGATG	TTATTAGAAA	2880
AGAAAATGCT	GACAAGGATG	AAGATGAAAA	AGGCGCAGAT	GAAGAAAGAC	ATAGTACTTC	2940
TGAAAGCTTA	AGTTCACCTG	AAGAAAAAAT	GTTAAGTATG	AATGAAGGAG	GAAATAGTTT	3000
AAATCATGAA	GAGGTGAAAG	AACATACTAG	TAATTCCTGAT	AATGTTCAAC	AGTCTGGAGG	3060
AATTGTTAAT	ATGAATGTTG	AGAAAGAACT	AAAAGATACT	TTAGAAAATC	CTTCTAGTAG	3120
CTTGGATGAA	GGAAAAGCAC	ATGAAGAATT	ATCAGAACCA	AATCTAAGCA	GTGACCAAGA	3180
TATGTCTAAT	ACACCTGGAC	CTTTGGATAA	CACCAGTGAA	GAAACTACAG	AAAGAATTAG	3240
TAATAATGAA	TATAAAGTTA	ACGAGAGGGA	AGATGAGAGA	ACGCTTACTA	AGGAATATGA	3300
AGATATTGTT	TTGAAAAGTC	ATATGAATAG	AGAATCAGAC	GATGGTGAAT	TATATGACGA	3360
AAATTCAGAC	TTATCTACTG	TAAATGATGA	ATCAGAAGAC	GCTGAAGCAA	AAATGAAAGG	3420
AAATGATACA	TCTGAAATGT	CGCATAATAG	TAGTCAACAT	ATTGAGAGTG	ATCAACAGAA	3480
AAACGATATG	AAAACCTGTTG	GTGATTTGGG	AACCACACAT	GTACAAAACG	AAATTAGTGT	3540
TCCTGTTACA	GGAGAAATTG	ATGAAAAAAT	AAGGGAAAGT	AAAGAATCAA	AAATTCATAA	3600
GGCTGAAGAG	GAAAGATTAA	GTCATACAGA	TATACATAAA	ATTAATCCTG	AAGATAGAAA	3660
TAGTAATACA	TTACATTTAA	AAGATATAAG	AAATGAGGAA	AACGAAAGAC	ACTTAACTAA	3720
TCAAAACATT	AATATTAGTC	AAGAAAGGGA	TTTGCAAAAA	CATGGATTCC	ATACCATGAA	3780
TAATCTACAT	GGAGATGGAG	TTTCCGAAAG	AAGTCAAATT	AATCATAGTC	ATCATGGAAA	3840
CAGACAAGAT	CGGGGGGGAA	ATTCTGGGAA	TGTTTTAAAT	ATGAGATCTA	ATAATAATAA	3900
TTTTAATAAT	ATTCCAAGTA	GATATAATTT	ATATGATAAA	AAATTAGATT	TAGATCTTTA	3960
TGAAAACAGA	AATGATAGTA	CAACAAAAGA	ATTAATAAAG	AAATTAGCAG	AAATAAATAA	4020
ATGTGAGAAC	GAAATTTCTG	TAAATATTG	TGACCATATG	ATTCATGAAG	AAATCCCATT	4080
AAAAACATGC	ACTAAAGAAA	AAACAAGAAA	TCTGTGTTGT	GCAGTATCAG	ATTACTGTAT	4140
GAGCTATTTT	ACATATGATT	CAGAGGAATA	TTATAATTGT	ACGAAAAGGG	AATTTGATGA	4200
TCCATCTTAT	ACATGTTTCA	GAAAGGAGGC	TTTTTCAAGT	ATGATATTCA	AATTTTTAAT	4260
AACAAATAAA	ATATATTATT	ATTTTTATAC	TTACAAAAC	GCAAAAGTAA	CAATAAAAAA	4320
AATTAATTC	TCATTAATTT	TTTTTTTCTT	TTTTTCTTTT	TAGGTATGCC	ATATTATGCA	4380
GGAGCAGGTG	TGTTATTTAT	TATATTGGTT	ATTTTAGGTG	CTTCACAAGC	CAAATATCAA	4440
AGGTTAGAAA	AAATAAATAA	AAATAAAATT	GAGAAGAATG	TAAATTAAAT	ATAGAATTCG	4500
AGCTCGG						4507

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1426 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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Met Lys Cys Asn Ile Ser Ile Tyr Phe Phe Ala Ser Phe Phe Val Leu
 1           5           10           15
Tyr Phe Ala Lys Ala Arg Asn Glu Tyr Asp Ile Lys Glu Asn Glu Lys
          20           25           30
Phe Leu Asp Val Tyr Lys Glu Lys Phe Asn Glu Leu Asp Lys Lys Lys
          35           40           45
Tyr Gly Asn Val Gln Lys Thr Asp Lys Lys Ile Phe Thr Phe Ile Glu
          50           55           60
Asn Lys Leu Asp Ile Leu Asn Asn Ser Lys Phe Asn Lys Arg Trp Lys
          65           70           75           80
Ser Tyr Gly Thr Pro Asp Asn Ile Asp Lys Asn Met Ser Leu Ile Asn
          85           90           95
Lys His Asn Asn Glu Glu Met Phe Asn Asn Asn Tyr Gln Ser Phe Leu
          100          105          110
Ser Thr Ser Ser Leu Ile Lys Gln Asn Lys Tyr Val Pro Ile Asn Ala
          115          120          125
Val Arg Val Ser Arg Ile Leu Ser Phe Leu Asp Ser Arg Ile Asn Asn
          130          135          140
Gly Arg Asn Thr Ser Ser Asn Asn Glu Val Leu Ser Asn Cys Arg Glu
          145          150          155          160
Lys Arg Lys Gly Met Lys Trp Asp Cys Lys Lys Lys Asn Asp Arg Ser
          165          170          175
Asn Tyr Val Cys Ile Pro Asp Arg Arg Ile Gln Leu Cys Ile Val Asn
          180          185          190
Leu Ser Ile Ile Lys Thr Tyr Thr Lys Glu Thr Met Lys Asp His Phe
          195          200          205
Ile Glu Ala Ser Lys Lys Glu Ser Gln Leu Phe Ser Leu Lys Asn Asp
          210          215          220
Asn Lys Tyr Asn Ser Lys Phe Cys Asn Asp Leu Lys Asn Ser Phe Leu
          225          230          235          240
Asp Tyr Gly His Leu Ala Met Gly Asn Asp Met Asp Phe Gly Gly Tyr
          245          250          255

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Ser Thr Lys Ala Glu Asn Lys Ile Gln Glu Val Phe Lys Gly Ala His
 260 265 270
 Gly Glu Ile Ser Glu His Lys Ile Lys Asn Ph Arg Lys Glu Trp Trp
 275 280 285
 Asn Glu Phe Arg Glu Lys Leu Trp Glu Ala Met Leu Ser Glu His Lys
 290 295 300
 Asn Asn Ile Asn Asn Cys Lys Asn Ile Ser Gln Glu Glu Leu Gln Ile
 305 310 315 320
 Thr Gln Trp Ile Lys Glu Trp His Gly Glu Phe Leu Leu Glu Arg Asp
 325 330 335
 Asn Arg Ser Lys Leu Pro Lys Ser Lys Cys Lys Asn Asn Thr Leu Tyr
 340 345 350
 Glu Ala Cys Glu Lys Glu Cys Ile Asp Pro Cys Met Lys Tyr Arg Asp
 355 360 365
 Trp Ile Ile Arg Ser Lys Phe Glu Trp His Thr Leu Ser Lys Glu Tyr
 370 375 380
 Glu Thr Gln Lys Val Pro Lys Glu Asn Ala Glu Asn Tyr Leu Ile Lys
 385 390 395 400
 Ile Ser Glu Asn Lys Asn Asp Ala Lys Val Ser Leu Leu Leu Asn Asn
 405 410 415
 Cys Asp Ala Glu Tyr Ser Lys Tyr Cys Asp Cys Lys His Thr Thr Thr
 420 425 430
 Leu Val Lys Ser Val Leu Asn Gly Asn Asp Asn Thr Ile Lys Glu Lys
 435 440 445
 Arg Glu His Ile Asp Leu Asp Asp Phe Ser Lys Phe Gly Cys Asp Lys
 450 455 460
 Asn Ser Val Asp Thr Asn Thr Lys Val Trp Glu Cys Lys Asn Pro Tyr
 465 470 475 480
 Ile Leu Ser Thr Lys Asp Val Cys Val Pro Pro Arg Arg Gln Glu Leu
 485 490 495
 Cys Leu Gly Asn Ile Asp Arg Ile Tyr Asp Lys Asn Leu Leu Met Ile
 500 505 510
 Lys Glu His Ile Leu Ala Ile Ala Ile Tyr Glu Ser Arg Ile Leu Lys
 515 520 525
 Arg Lys Tyr Lys Asn Lys Asp Asp Lys Glu Val Cys Lys Ile Ile Asn
 530 535 540
 Lys Thr Phe Ala Asp Ile Arg Asp Ile Ile Gly Gly Thr Asp Tyr Trp
 545 550 555 560
 Asn Asp Leu Ser Asn Arg Lys Leu Val Gly Lys Ile Asn Thr Asn Ser
 565 570 575
 Lys Tyr Val His Arg Asn Lys Lys Asn Asp Lys Leu Phe Arg Asp Glu
 580 585 590
 Trp Trp Lys Val Il Lys Lys Asp Val Trp Asn Val Ile Ser Trp Val
 595 600 605

Phe Lys Asp Lys Thr Val Cys Lys Glu Asp Asp Ile Glu Asn Ile Pro
 610 615 620
 Gln Phe Phe Arg Trp Phe Ser Glu Trp Gly Asp Asp Tyr Cys Gln Asp
 625 630 635 640
 Lys Thr Lys Met Ile Glu Thr Leu Lys Val Glu Cys Lys Glu Lys Pro
 645 650 655
 Cys Glu Asp Asp Asn Cys Lys Ser Lys Cys Asn Ser Tyr Lys Glu Trp
 660 665 670
 Ile Ser Lys Lys Lys Lys Ser Ile Ile Thr Ser Asn Ile Pro Arg Ile
 675 680 685
 Ser Lys Arg Asn Asn Tyr Lys Met Tyr Ser Glu Phe Lys Ser Ile Lys
 690 695 700
 Pro Glu Val Tyr Leu Lys Lys Tyr Ser Lys Cys Ser Asn Leu Asn Phe
 705 710 715 720
 Glu Asp Glu Phe Lys Glu Glu Leu His Ser Asp Tyr Lys Asn Lys Cys
 725 730 735
 Thr Met Cys Pro Glu Val Lys Asp Val Pro Ile Ser Ile Ile Arg Asn
 740 745 750
 Asn Glu Gln Thr Ser Gln Glu Ala Val Pro Glu Glu Asn Thr Glu Ile
 755 760 765
 Ala His Arg Thr Glu Thr Pro Ser Ile Ser Glu Gly Pro Lys Gly Asn
 770 775 780
 Glu Gln Lys Glu Arg Asp Asp Ser Leu Ser Lys Ile Ser Val Ser
 785 790 795 800
 Pro Glu Asn Ser Arg Pro Glu Thr Asp Ala Lys Asp Thr Ser Asn Leu
 805 810 815
 Leu Lys Leu Lys Gly Asp Val Asp Ile Ser Met Pro Lys Ala Val Ile
 820 825 830
 Gly Ser Ser Pro Asn Asp Asn Ile Asn Val Thr Glu Gln Gly Asp Asn
 835 840 845
 Ile Ser Gly Val Asn Ser Lys Pro Leu Ser Asp Asp Val Arg Pro Asp
 850 855 860
 Lys Lys Glu Leu Glu Asp Gln Asn Ser Asp Glu Ser Glu Glu Thr Val
 865 870 875 880
 Val Asn His Ile Ser Lys Ser Pro Ser Ile Asn Asn Gly Asp Asp Ser
 885 890 895
 Gly Ser Gly Ser Ala Thr Val Ser Glu Ser Ser Ser Ser Asn Thr Gly
 900 905 910
 Leu Ser Ile Asp Asp Asp Arg Asn Gly Asp Thr Phe Val Arg Thr Gln
 915 920 925
 Asp Thr Ala Asn Thr Glu Asp Val Ile Arg Lys Glu Asn Ala Asp Lys
 930 935 940
 Asp Glu Asp Glu Lys Gly Ala Asp Glu Glu Arg His Ser Thr Ser Glu
 945 950 955 960

Ser Leu Ser Ser Pro Glu Glu Lys Met Leu Thr Asp Asn Glu Gly Gly
 965 970 975
 Asn Ser Leu Asn His Glu Glu Val Lys Glu His Thr Ser Asn Ser Asp
 980 985 990
 Asn Val Gln Gln Ser Gly Gly Ile Val Asn Met Asn Val Glu Lys Glu
 995 1000 1005
 Leu Lys Asp Thr Leu Glu Asn Pro Ser Ser Ser Leu Asp Glu Gly Lys
 1010 1015 1020
 Ala His Glu Glu Leu Ser Glu Pro Asn Leu Ser Ser Asp Gln Asp Met
 1025 1030 1035 1040
 Ser Asn Thr Pro Gly Pro Leu Asp Asn Thr Ser Glu Glu Thr Thr Glu
 1045 1050 1055
 Arg Ile Ser Asn Asn Glu Tyr Lys Val Asn Glu Arg Glu Asp Glu Arg
 1060 1065 1070
 Thr Leu Thr Lys Glu Tyr Glu Asp Ile Val Leu Lys Ser His Met Asn
 1075 1080 1085
 Arg Glu Ser Asp Asp Gly Glu Leu Tyr Asp Glu Asn Ser Asp Leu Ser
 1090 1095 1100
 Thr Val Asn Asp Glu Ser Glu Asp Ala Glu Ala Lys Met Lys Gly Asn
 1105 1110 1115 1120
 Asp Thr Ser Glu Met Ser His Asn Ser Ser Gln His Ile Glu Ser Asp
 1125 1130 1135
 Gln Gln Lys Asn Asp Met Lys Thr Val Gly Asp Leu Gly Thr Thr His
 1140 1145 1150
 Val Gln Asn Glu Ile Ser Val Pro Val Thr Gly Glu Ile Asp Glu Lys
 1155 1160 1165
 Leu Arg Glu Ser Lys Glu Ser Lys Ile His Lys Ala Glu Glu Glu Arg
 1170 1175 1180
 Leu Ser His Thr Asp Ile His Lys Ile Asn Pro Glu Asp Arg Asn Ser
 1185 1190 1195 1200
 Asn Thr Leu His Leu Lys Asp Ile Arg Asn Glu Glu Asn Glu Arg His
 1205 1210 1215
 Leu Thr Asn Gln Asn Ile Asn Ile Ser Gln Glu Arg Asp Leu Gln Lys
 1220 1225 1230
 Ala Val Asp Ser Met Glu Met Glu Phe Pro Lys Glu Val Lys Leu Ile
 1235 1240 1245
 Ile Val Ile Met Glu Thr Asp Lys Ile Gly Gly Asn Ser Gly Asn Val
 1250 1255 1260
 Leu Asn Met Arg Ser Asn Asn Asn Asn Phe Asn Asn Ile Pro Ser Arg
 1265 1270 1275 1280
 Tyr Asn Leu Tyr Asp Lys Lys Leu Asp Leu Asp Leu Tyr Glu Asn Arg
 1285 1290 1295
 Asn Asp Ser Thr Thr Lys Glu Leu Ile Lys Lys Leu Ala Glu Ile Asn
 1300 1305 1310

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Lys Cys Glu Asn Glu Ile Ser Val Lys Tyr Cys Asp His Met Ile His
1315 1320 1325

Glu Glu Il Pro Leu Lys Thr Cys Thr Lys Glu Lys Thr Arg Asn Leu
1330 1335 1340

Cys Cys Ala Val Ser Asp Tyr Cys Met Ser Tyr Phe Thr Tyr Asp Ser
1345 1350 1355 1360

Glu Glu Tyr Tyr Asn Cys Thr Lys Arg Glu Phe Asp Asp Pro Ser Tyr
1365 1370 1375

Thr Cys Phe Arg Lys Glu Ala Phe Ser Ser Met Ile Phe Lys Phe Leu
1380 1385 1390

Ile Thr Asn Lys Ile Tyr Tyr Tyr Phe Tyr Thr Tyr Lys Thr Ala Lys
1395 1400 1405

Val Thr Ile Lys Lys Ile Asn Phe Ser Leu Ile Phe Phe Phe Phe
1410 1415 1420

Ser Phe
1425

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2288 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CACTTTATGC TTCCGGCTCG TATGTTGTGT GGAATTGTGA GCGGATAACA ATTCACACA	60
GGAAACAGCT ATGACCATGA TTACGCCAAG CTCTAATACG ACTCACTATA GGGAAAGCTG	120
GTACGCCTGC AGGTCCGGTC CGGAATTCAA TAAATATTT CCAGAAAGGA ATGTGCAAAT	180
TCACATATCC AATATATTCA AGGAATATAA AGAAAATAAT GTAGATATCA TATTGGAAC	240
GTTGAATTAT GAATATAATA ATTTCTGTAA AGAAAAACCT GAATTAGTAT CTGCTGCCAA	300
GTATAATCTG AAAGCTCCAA ATGCTAAATC CCCTAGAATA TACAAATCTA AGGAGCATGA	360
AGAATCAAGT GTGTTTGGTT GCAAAACGAA AATCAGTAAA GTTAAAAAAA AATGGAATTG	420
TTATAGTAAT AATAAAGTAA CTAAACCTGA AGGTGTATGT GGACCACCAA GAAGGCAACA	480
ATTATGTCTT GGATATATAT TTTTGATTCTG CGACGGTAAC GAGGAAGGAT TAAAAGATCA	540
TATTAATAAG GCAGCTAATT ATGAGGCAAT GCATTTAAAA GAGAAATATG AGAATGCTGG	600
TGGTGATAAA ATTTGCAATG CTATATTGGG AAGTTATGCA GATATTGGAG ATATTGTAAG	660
AGGTTTGGAT GTTTGGAGGG ATATAAATAC TAATAAATTA TCAGAAAAAT TCCAAAAAAT	720
TTTTATGGGT GGTGGTAATT CTAGGAAAAA ACAAACGAT AATAATGAAC GTAATAAATG	780
GTGGGAAAAA CAAAGGAATT TAATATGGTC TAGTATGGTA AAACACATTC CAAAGGAAA	840
AACATGTAAA CGTCATAATA ATTTTGAGAA AATTCCTCAA TTTTGGAGAT GGTAAAAGA	900
ATGGGGTGAT GAATTTTGTG AGGAAATGGG TACGGAAGTC AAGCAATTAG AGAAAATATG	960
TGAAAATAAA AATTGTTCTG AAAAAAATG TAAAAATGCA TGAGTTCCT ATGAAAATG	1020
GATAAAGGAA CGAAAAATG AATATAATTT GCAATCAAAG AAATTTGATA GTGATAAAAA	1080
ATTAAATAAA AAAACAATC TTTATAATAA ATTTGAGGAT TCTAAAGCTT ATTTAAGGAG	1140
TGAATCAAAA CAGTGCTCAA ATATAGAATT TAATGATGAA ACATTTACAT TTCCTAATAA	1200
ATATAAAGAG GCTTGATGG TATGTGAAAA TCCTTCATCT TCGAAAGCTC TTAAACCTAT	1260
AAAAACGAAT GTGTTTCCTA TAGAGGAATC AAAAAAATCT GAGTTATCAA GTTTAACAGA	1320
TAAATCTAAG AATACTCCTA ATAGTTCTGG TGGGGGAAAT TATGGAGATA GACAAATATC	1380
AAAAAGAGAC GATGTTTCATC ATGATGGTCC TAAGGAAGTG AAATCCGGAG AAAAGAGGT	1440
ACCAAAAATA GATGCAGCTG TTAAAACAGA AAATGAATTT ACCTCTAATC GAAACGATAT	1500

TGAAGGAAAG GAAAAAAGTA AAGGTGATCA TTCTTCTCCT GTTCATTCTA AAGATATAAA 1560
 AAATGAGGAA CCACAAAGGG TGGTGTCTGA AAATTTACCT AAAATTGAAG AGAAAATGGA 1620
 ATCTTCTGAT TCTATACCAA TTAATCATAT AGAAGCTGAA AAGGGTCAGT CTTCTAATTC 1680
 TAGCGATAAT GATCCTGCAG TAGTAAGTGG TAGAGAATCT AAAGATGTAA ATCTTCATAC 1740
 TTCTGAAAGG ATTAAAGAAA ATGAAGAAGG TGTGATTAAA ACAGATGATA GTTCAAAAAG 1800
 TATTGAAATT TCTAAAATAC CATCTGACCA AAATAATCAT AGTGATTAT CACAGAATGC 1860
 AAATGAGGAC TCTAATCAAG GGAATAAGGA AACAATAAAT CCTCCTTCTA CAGAAAAAAA 1920
 TCTCAAAGAA ATTCATTATA AAACATCTGA TTCTGATGAT CATGGTTCTA AAATTAAAAG 1980
 TGAAATTGAA CCAAAGGAGT TAACGGAGGA ATCACCTCTT ACTGATAAAA AAAGTGAAG 2040
 TGCAGCGATT GGTGATAAAA ATCATGAATC AGTAAAAGC GCTGATATTT TTCAATCTGA 2100
 GATTCATAAT TCTGATAATA GAGATAGAAT TGTTTCTGAA AGTGTAGTTC AGGATTCTTC 2160
 AGGAAGCTCT ATGAGTACTG AATCTATACG TACTGATAAC AAGGATTTTA AAACAAGTGA 2220
 GGATATTGCA CCTTCTATTA ATGGTCGGAA TTCCCGGGTC GACGAGCTCA CTAGTCGGCG 2280
 GCCGCTCT 2288

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 749 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Ala Asp Asn Asn Phe Thr Gln Glu Thr Ala Met Thr Met Ile Thr Pro
 1 5 10 15
 Ser Ser Asn Thr Thr His Tyr Arg Glu Ser Trp Tyr Ala Cys Arg Ser
 20 25 30
 Gly Pro Glu Phe Asn Lys Ile Phe Pro Glu Arg Asn Val Gln Ile His
 35 40 45
 Ile Ser Asn Ile Phe Lys Glu Tyr Lys Glu Asn Asn Val Asp Ile Ile
 50 55 60
 Phe Gly Thr Leu Asn Tyr Glu Tyr Asn Asn Phe Cys Lys Glu Lys Pro
 65 70 75 80
 Glu Leu Val Ser Ala Ala Lys Tyr Asn Leu Lys Ala Pro Asn Ala Lys
 85 90 95
 Ser Pro Arg Ile Tyr Lys Ser Lys Glu His Glu Glu Ser Ser Val Ph
 100 105 110

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Gly Cys Lys Thr Lys Ile Ser Lys Val Lys Lys Lys Trp Asn Cys Tyr
 115 120 125
 Ser Asn Asn Lys Val Thr Lys Pro Glu Gly Val Cys Gly Pro Pro Arg
 130 135 140
 Arg Gln Gln Leu Cys Leu Gly Tyr Ile Phe Leu Ile Arg Asp Gly Asn
 145 150 155 160
 Glu Glu Gly Leu Lys Asp His Ile Asn Lys Ala Ala Asn Tyr Glu Ala
 165 170 175
 Met His Leu Lys Glu Lys Tyr Glu Asn Ala Gly Gly Asp Lys Ile Cys
 180 185 190
 Asn Ala Ile Leu Gly Ser Tyr Ala Asp Ile Gly Asp Ile Val Arg Gly
 195 200 205
 Leu Asp Val Trp Arg Asp Ile Asn Thr Asn Lys Leu Ser Glu Lys Phe
 210 215 220
 Gln Lys Ile Phe Met Gly Gly Gly Asn Ser Arg Lys Lys Gln Asn Asp
 225 230 235 240
 Asn Asn Glu Arg Asn Lys Trp Trp Glu Lys Gln Arg Asn Leu Ile Trp
 245 250 255
 Ser Ser Met Val Lys His Ile Pro Lys Gly Lys Thr Cys Lys Arg His
 260 265 270
 Asn Asn Phe Glu Lys Ile Pro Gln Phe Leu Arg Trp Leu Lys Glu Trp
 275 280 285
 Gly Asp Glu Phe Cys Glu Glu Met Gly Thr Glu Val Lys Gln Leu Glu
 290 295 300
 Lys Ile Cys Glu Asn Lys Asn Cys Ser Glu Lys Lys Cys Lys Asn Ala
 305 310 315 320
 Cys Ser Ser Tyr Glu Lys Trp Ile Lys Glu Arg Lys Asn Glu Tyr Asn
 325 330 335
 Leu Gln Ser Lys Lys Phe Asp Ser Asp Lys Lys Leu Asn Lys Lys Asn
 340 345 350
 Asn Leu Tyr Asn Lys Phe Glu Asp Ser Lys Ala Tyr Leu Arg Ser Glu
 355 360 365
 Ser Lys Gln Cys Ser Asn Ile Glu Phe Asn Asp Glu Thr Phe Thr Phe
 370 375 380
 Pro Asn Lys Tyr Lys Glu Ala Cys Met Val Cys Glu Asn Pro Ser Ser
 385 390 395 400
 Ser Lys Ala Leu Lys Pro Ile Lys Thr Asn Val Phe Pro Ile Glu Glu
 405 410 415
 Ser Lys Lys Ser Glu Leu Ser Ser Leu Thr Asp Lys Ser Lys Asn Thr
 420 425 430
 Pro Asn Ser Ser Gly Gly Gly Asn Tyr Gly Asp Arg Gln Ile Ser Lys
 435 440 445
 Arg Asp Asp Val His His Asp Gly Pro Lys Glu Val Lys Ser Gly Glu
 450 455 460

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Lys Glu Val Pro Lys Ile Asp Ala Ala Val Lys Thr Glu Asn Glu Phe
 465 470 475 480
 Thr Ser Asn Arg Asn Asp Ile Glu Gly Lys Glu Lys S r Lys Gly Asp
 485 490 495
 His Ser Ser Pro Val His Ser Lys Asp Ile Lys Asn Glu Glu Pro Gln
 500 505 510
 Arg Val Val Ser Glu Asn Leu Pro Lys Ile Glu Glu Lys Met Glu Ser
 515 520 525
 Ser Asp Ser Ile Pro Ile Thr His Ile Glu Ala Glu Lys Gly Gln Ser
 530 535 540
 Ser Asn Ser Ser Asp Asn Asp Pro Ala Val Val Ser Gly Arg Glu Ser
 545 550 555 560
 Lys Asp Val Asn Leu His Thr Ser Glu Arg Ile Lys Glu Asn Glu Glu
 565 570 575
 Gly Val Ile Lys Thr Asp Asp Ser Ser Lys Ser Ile Glu Ile Ser Lys
 580 585 590
 Ile Pro Ser Asp Gln Asn Asn His Ser Asp Leu Ser Gln Asn Ala Asn
 595 600 605
 Glu Asp Ser Asn Gln Gly Asn Lys Glu Thr Ile Asn Pro Pro Ser Thr
 610 615 620
 Glu Lys Asn Leu Lys Glu Ile His Tyr Lys Thr Ser Asp Ser Asp Asp
 625 630 635 640
 His Gly Ser Lys Ile Lys Ser Glu Ile Glu Pro Lys Glu Leu Thr Glu
 645 650 655
 Glu Ser Pro Leu Thr Asp Lys Lys Thr Glu Ser Ala Ala Ile Gly Asp
 660 665 670
 Lys Asn His Glu Ser Val Lys Ser Ala Asp Ile Phe Gln Ser Glu Ile
 675 680 685
 His Asn Ser Asp Asn Arg Asp Arg Ile Val Ser Glu Ser Val Val Gln
 690 695 700
 Asp Ser Ser Gly Ser Ser Met Ser Thr Glu Ser Ile Arg Thr Asp Asn
 705 710 715 720
 Lys Asp Phe Lys Thr Ser Glu Asp Ile Ala Pro Ser Ile Asn Gly Arg
 725 730 735
 Asn Ser Arg Val Asp Glu Leu Thr Ser Arg Arg Pro Leu
 740 745

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2606 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Plasmodium falciparum*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

AGCTCTATTA CGACTCACTA TAGGGAAAGC TGGTACGCCT GCAGGTACCG GTCCGGAATT	60
CCCGGGTCTGA CGAGCTCACT AGTCGGCGGC CGCTCTAGAG GATCCAAGCT TAATAGTGTT	120
TATACGTCTA TTGGCTTATT TTAAATAGC TAAAAAGCG GACCATGTAA AAAGGATAAT	180
GATAATGCAG AGGATAATAT AGATTTTGGT GATGAAGGTA AAACATTTAA AGAGGCAGAT	240
AATTGTAAAC CATGTTCTCA ATTTACTGTT GATTGTAAAA ATTGTAATGG TGGTGATACA	300
AAAGGGAAGT GCAATGGCAG CAATGGCAA AAGAATGGAA ATGATTATAT TACTGCAAGT	360
GATATTGAAA ATGGAGGGAA TTCTATTGGA AATATAGATA TGGTTGTTAG TGATAAGGAT	420
GCAAATGGAT TTAATGGTTT AGACGCTTGT GGAAGTGCAA ATATCTTTAA AGGTATTAGA	480
AAAGAACAAT GGAAATGTGC TAAAGTATGT GGTTTAGATG TATGTGGTCT TAAAAATGGT	540
AATGGTAGTA TAGATAAAGA TCAAAAACAA ATTATAATTA TTAGAGCATT GCTTAAACGT	600
TGGGTAGAAT ATTTTTTAGA AGATTATAAT AAAATTAATG CCAAATTTT ACATTGTACG	660
AAAAAGGATA ATGAATCCAC ATGTACAAAT GATTGTCCAA ATAAATGTAC ATGTGTAGAA	720
GAGTGGATAA ATCAGAAAAG GACAGAATGG AAAAATATAA AAAACATTA CAAAACACAA	780
AATGAAAATG GTGACAATAA CATGAAATCT TTGGTTACAG ATATTTTGGG TGCCTTGCAA	840
CCCCAAAGTG ATGTTAACAA AGCTATAAAA CCTTGTTAGT GTTTAACTGC GTTCGAGAGT	900
TTTTGTGGTC TTAATGGCGC TGATAACTCA GAAAAAAAAG AAGGTGAAGA TTACGATCTT	960
GTTCTATGTA TGCTTAAAAA TCTTGAAAA CAAATTCAGG AGTGCAAAAA GAAACATGGC	1020
GAAACTAGTG TCGAAAATGG TGGCAAATCA TGTACCCCCC TTGACAACAC CACCCTTGAG	1080
GAGGAACCCA TAGAAGAGGA AAACCAAGTG GAAGCGCCGA ACATTGTGCC AAAACAAACA	1140
GTGGAAGATA AAAAAAAGA GGAAGAAGAA GAAACTTGTA CACCGGCATC ACCAGTACCA	1200
GAAAAACCGG TACCTCATGT GGCACGTTGG CGAACATTTA CACCACCTGA GGTATTCAAG	1260
ATATGGAGGG GAAGGAGAAA TAAACTACG TGCGAAATAG TGGCAGAAAT GCTTAAAGAT	1320
AAGAATGGAA GGAATACAGT AGGTGAATGT TATAGAAAAG AAACCTATTC TGAATGGACG	1380
TGTGATGAAA GTAAGATTAA AATGGGACAG CATGGAGCAT GTATTCCTCC AAGAAGACAA	1440
AAATTATGTT TACATTATTT AGAAAAATA ATGACAAATA CAAATGAATT GAAATACGCA	1500

TTTATTAAAT	GTGCTGCAGC	AGAAACTTTT	TTGTTATGGC	AAACTACAA	AAAAGATAAG	1560
AATGGTAATG	CAGAAGATCT	CGATGAAAAA	TTAAAAGGTG	GTATTATCCC	CGAAGATTTT	1620
AAACGGCAAA	TGTTCTATAC	GTTTGCAGAT	TATAGAGATA	TATGTTTGGG	TACGGATATA	1680
TCATCAAAAA	AAGATACAAG	TAAAGGTGTA	GGTAAAGTAA	AATGCAATAT	TGATGATGTT	1740
TTTTATAAAA	TTAGCAATAG	TATTCGTTAC	CGTAAAAGTT	GGTGGGAAAC	AAATGGTCCA	1800
GTTATATGGG	AAGGAATGTT	ATGCGCTTTA	AGTTATGATA	CGAGCCTAAA	TAATGTTAAT	1860
CCGGAAGCTC	ACAAAAAACT	TACCGAAGGC	AATAACAACT	TTGAGAAAGT	CATATTTGGT	1920
AGTGATAGTA	GCACTACTTT	GTCCAAATTT	TCTGAAAGAC	CTCAATTTCT	AAGATGGTTG	1980
ACTGAATGGG	GAGAAAATTT	CTGCAAAGAA	CAAAAAAAGG	AGTATAAGGT	GTTGTTGGCA	2040
AAATGTAAGG	ATTGTGATGT	TGATGGTGAT	GGTAAATGTA	ATGGAAAATG	TGTTGCGTGC	2100
AAAGATCAAT	GTAAACAATA	TCATAGTTGG	ATTGGAATAT	GGATAGATAA	TTATAAAAAA	2160
CAAAAAGGAA	GATATACTGA	GGTTAAAAAA	ATACCTCTGT	ATAAGAAGA	TAAAGACGTG	2220
AAAAACTCAG	ATGATGCTCG	CGATTATTTA	AAAACACAAT	TACAAAATAT	GAAATGTGTA	2280
AATGGAACCTA	CTGATGAAAA	TTGTGAGTAT	AAGTGTATGC	ATAAACCTC	ATCCACAAAT	2340
AGTGATATGC	CCGAATCGTT	GGACGAAAAG	CCGGAAAAGG	TCAAAGACAA	GTGTAATTGT	2400
GTACCTAATG	AATGCAATGC	ATTGAGTGTA	AGTGGTAGCG	GTTTTCTCTGA	TGGTCAAGCT	2460
TACGTACGCG	TGCATGCGAC	GTCATAGCTC	TTCTATAGTG	TCACCTAAAT	TCAATTCACT	2520
GGCCGTCGTT	TTACAACGTC	GTGACTGGGA	AAACCTGGCG	TTACCCAACCT	TAATCGCCTT	2580
GCAGCACATC	CCCCTTTTCGC	CAGCTG				2606

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 921 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

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Lys Leu Asn Ser Val Tyr Thr Ser Ile Gly Leu Phe Leu Asn Ser Leu
1           5           10           15
Lys Ser Gly Pro Cys Lys Lys Asp Asn Asp Asn Ala Glu Asp Asn Ile
          20           25           30
Asp Phe Gly Asp Glu Gly Lys Thr Phe Lys Glu Ala Asp Asn Cys Lys
          35           40           45
Pro Cys Ser Gln Phe Thr Val Asp Cys Lys Asn Cys Asn Gly Gly Asp
          50           55           60
Thr Lys Gly Lys Cys Asn Gly Ser Asn Gly Lys Lys Asn Gly Asn Asp
          65           70           75           80
Tyr Ile Thr Ala Ser Asp Ile Glu Asn Gly Gly Asn Ser Ile Gly Asn
          85           90           95
Ile Asp Met Val Val Ser Asp Lys Asp Ala Asn Gly Phe Asn Gly Leu
          100          105          110
Asp Ala Cys Gly Ser Ala Asn Ile Phe Lys Gly Ile Arg Lys Glu Gln
          115          120          125
Trp Lys Cys Ala Lys Val Cys Gly Leu Asp Val Cys Gly Leu Lys Asn
          130          135          140
Gly Asn Gly Ser Ile Asp Lys Asp Gln Lys Gln Ile Ile Ile Ile Arg
          145          150          155          160
Ala Leu Leu Lys Arg Trp Val Glu Tyr Phe Leu Glu Asp Tyr Asn Lys
          165          170          175
Ile Asn Ala Lys Ile Ser His Cys Thr Lys Lys Asp Asn Glu Ser Thr
          180          185          190
Cys Thr Asn Asp Cys Pro Asn Lys Cys Thr Cys Val Glu Glu Trp Ile
          195          200          205
Asn Gln Lys Arg Thr Glu Trp Lys Asn Ile Lys Lys His Tyr Lys Thr
          210          215          220
Gln Asn Glu Asn Gly Asp Asn Asn Met Lys Ser Leu Val Thr Asp Ile
          225          230          235          240
Leu Gly Ala Leu Gln Pro Gln Ser Asp Val Asn Lys Ala Il Lys Pro
          245          250          255

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Cys Ser Gly Leu Thr Ala Phe Glu Ser Phe Cys Gly Leu Asn Gly Ala
 260 265 270
 Asp Asn Ser Glu Lys Lys Glu Gly Glu Asp Tyr Asp Leu Val Leu Cys
 275 280 285
 Met Leu Lys Asn Leu Glu Lys Gln Ile Gln Glu Cys Lys Lys Lys His
 290 295 300
 Gly Glu Thr Ser Val Glu Asn Gly Gly Lys Ser Cys Thr Pro Leu Asp
 305 310 315 320
 Asn Thr Thr Leu Glu Glu Glu Pro Ile Glu Glu Glu Asn Gln Val Glu
 325 330 335
 Ala Pro Asn Ile Cys Pro Lys Gln Thr Val Glu Asp Lys Lys Lys Glu
 340 345 350
 Glu Glu Glu Glu Thr Cys Thr Pro Ala Ser Pro Val Pro Glu Lys Pro
 355 360 365
 Val Pro His Val Ala Arg Trp Arg Thr Phe Thr Pro Pro Glu Val Phe
 370 375 380
 Lys Ile Trp Arg Gly Arg Arg Asn Lys Thr Thr Cys Glu Ile Val Ala
 385 390 395 400
 Glu Met Leu Lys Asp Lys Asn Gly Arg Thr Thr Val Gly Glu Cys Tyr
 405 410 415
 Arg Lys Glu Thr Tyr Ser Glu Trp Thr Cys Asp Glu Ser Lys Ile Lys
 420 425 430
 Met Gly Gln His Gly Ala Cys Ile Pro Pro Arg Arg Gln Lys Leu Cys
 435 440 445
 Leu His Tyr Leu Glu Lys Ile Met Thr Asn Thr Asn Glu Leu Lys Tyr
 450 455 460
 Ala Phe Ile Lys Cys Ala Ala Ala Glu Thr Phe Leu Leu Trp Gln Asn
 465 470 475 480
 Tyr Lys Lys Asp Lys Asn Gly Asn Ala Glu Asp Leu Asp Glu Lys Leu
 485 490 495
 Lys Gly Gly Ile Ile Pro Glu Asp Phe Lys Arg Gln Met Phe Tyr Thr
 500 505 510
 Phe Ala Asp Tyr Arg Asp Ile Cys Leu Gly Thr Asp Ile Ser Ser Lys
 515 520 525
 Lys Asp Thr Ser Lys Gly Val Gly Lys Val Lys Cys Asn Ile Asp Asp
 530 535 540
 Val Phe Tyr Lys Ile Ser Asn Ser Ile Arg Tyr Arg Lys Ser Trp Trp
 545 550 555 560
 Glu Thr Asn Gly Pro Val Ile Trp Glu Gly Met Leu Cys Ala Leu Ser
 565 570 575
 Tyr Asp Thr Ser Leu Asn Asn Val Asn Pro Glu Thr His Lys Lys Leu
 580 585 590
 Thr Glu Gly Asn Asn Asn Phe Glu Lys Val Ile Phe Gly Ser Asp Ser
 595 600 605

55

Ser Thr Thr Leu Ser Lys Phe Ser Glu Arg Pro Gln Phe Leu Arg Trp
 610 615 620
 Leu Thr Glu Trp Gly Glu Asn Phe Cys Lys Glu Gln Lys Lys Glu Tyr
 625 630 635 640
 Lys Val Leu Leu Ala Lys Cys Lys Asp Cys Asp Val Asp Gly Asp Gly
 645 650 655
 Lys Cys Asn Gly Lys Cys Val Ala Cys Lys Asp Gln Cys Lys Gln Tyr
 660 665 670
 His Ser Trp Ile Gly Ile Trp Ile Asp Asn Tyr Lys Lys Gln Lys Gly
 675 680 685
 Arg Tyr Thr Glu Val Lys Lys Ile Pro Leu Tyr Lys Glu Asp Lys Asp
 690 695 700
 Val Lys Asn Ser Asp Asp Ala Arg Asp Tyr Leu Lys Thr Gln Leu Gln
 705 710 715 720
 Asn Met Lys Cys Val Asn Gly Thr Thr Asp Glu Asn Cys Glu Tyr Lys
 725 730 735
 Cys Met His Lys Thr Ser Ser Thr Asn Ser Asp Met Pro Glu Ser Leu
 740 745 750
 Asp Glu Lys Pro Glu Lys Val Lys Asp Lys Cys Asn Cys Val Pro Asn
 755 760 765
 Glu Cys Asn Ala Leu Ser Val Ser Gly Ser Gly Phe Pro Asp Gly Gln
 770 775 780
 Ala Phe Gly Gly Gly Val Leu Glu Gly Thr Cys Lys Gly Leu Gly Glu
 785 790 795 800
 Pro Lys Lys Lys Ile Glu Pro Pro Gln Tyr Asp Pro Thr Asn Asp Ile
 805 810 815
 Leu Lys Ser Thr Ile Pro Val Thr Ile Val Leu Ala Leu Gly Ser Ile
 820 825 830
 Ala Phe Leu Phe Met Lys Val Ile Tyr Ile Tyr Val Trp Tyr Ile Tyr
 835 840 845
 Met Leu Cys Val Gly Ala Leu Asp Thr Tyr Ile Cys Gly Cys Ile Cys
 850 855 860
 Ile Cys Ile Phe Ile Cys Val Ser Val Tyr Val Cys Val Tyr Val Tyr
 865 870 875 880
 Val Phe Leu Tyr Met Cys Val Phe Tyr Ile Tyr Phe Ile Tyr Ile Tyr
 885 890 895
 Val Phe Ile Leu Lys Met Lys Lys Met Lys Lys Met Lys Lys Met Lys
 900 905 910
 Lys Met Lys Lys Arg Lys Lys Arg Ile
 915 920

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2101 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GGAACAGGGT GATAATAAAG TAGGAGCCTG TGCTCCGTAT AGACGATTAC ATTTATGTGA	60
TTATAATTTG GAATCTATAG ACACAACGTC GACGACGCAT AAGTTGTTGT TAGAGGTGTG	120
TATGGCAGCA AAATACGAAG GAAACTCAAT AAATACACAT TATACACAAC ATCAACGAAC	180
TAATGAGGAT TCTGCTTCCC AATTATGTAC TGTATTAGCA CGAAGTTTTG CAGATATAGG	240
TGATATCGTA AGAGGAAAAG ATCTATATCT CGGTTATGAT AATAAAGAAA AAGAACAAAG	300
AAAAAAATTA GAACAGAAAT TGAAAGATAT TTTCAAGAAA ATACATAAGG ACGTGATGAA	360
GACGAATGGC GCACAAGAAC GCTACATAGA TGATGCCAAA GGAGGAGATT TTTTCAATT	420
AAGAGAAGAT TGGTGGACGT CGAATCGAGA AACAGTATGG AAAGCATTAA TATGTCATGC	480
ACCAAAAGAA GCTAATTATT TTATAAAAAC AGCGTGTAAT GTAGGAAAAG GAACATAATGG	540
TCAATGCCAT TGCATTGGTG GAGATGTTCC CACATATTTT GATTATGTGC CGCAGTATCT	600
TCGCTGGTTC GAGGAATGGG CAGAAGACTT TTGCAGGAAA AAAAAAAAAA AACTAGAAAA	660
TTTGCAAAAA CAGTGTCGTG ATTACGAACA AAATTTATAT TGTAAGTGGTA ATGGCTACGA	720
TTGCACAAAA ACTATATATA AAAAAGGTAA ACTTGTTATA GGTGAACATT GTACAAACTG	780
TTCTGTTTGG TGTCGTATGT ATGAAACTTG GATAGATAAC CAGAAAAAAG AATTTCTAAA	840
ACAAAAAAGA AAATACGAAA CAGAAATATC AGGTGGTGGT AGTGGTAAGA GTCCTAAAAG	900
GACAAAACGG GCTGCACGTA GTAGTAGTAG TAGTGATGAT AATGGGTATG AAAGTAAATT	960
TTATAAAAAA CTGAAAGAAG TTGGCTACCA AGATGTCGAT AAATTTTTTA AAATATTAAA	1020
CAAAGAAGGA ATATGTCAAA AACAACTCA AGTAGGAAAT GAAAAGCAG ATAATGTTGA	1080
TTTTACTAAT GAAAAATATG TAAAAACATT TTCTCGTACA GAAATTTGTG AACCGTGCCC	1140
ATGGTGTGGA TTGGAAAAAG GTGGTCCACC ATGGAAAGTT AAAGGTGACA AAACCTGCGG	1200
AAGTGCAAAA ACAAGACAT ACGATCCTAA AAATATTACC GATATACCAG TACTCTACCC	1260
TGATAAATCA CAGCAAAATA TACTAAAAAA ATATAAAAAT TTTTGTGAAA AAGGTGCACC	1320
TGGTGGTGGT CAAATTAAAA AATGGCAATG TTATTATGAT GAACATAGGC CTAGTAGTAA	1380
AAATAATAAT AATTGTGTAG AAGGAACATG GGACAAGTTT ACACAAGGTA AACAAACCGT	1440
TAAGTCCTAT AATGTTTTTT TTTGGGATTG GGTCATGAT ATGTTACACG ATCTGTAGA	1500

GTGGAAGACA	GAACCTAGTA	AGTGTATAAA	TAATAACACT	AATGGCAACA	CATGTAGAAA	1560
CAATAATAAA	TGTAAAACAG	ATTGTGGTTG	TTTTCAAAAA	TGGGTTGAAA	AAAAACAACA	1620
AGAATGGATG	GCAATAAAAG	ACCATTTTGG	AAAGCAAACA	GATATTGTCC	AACAAAAAGG	1680
TCTTATCGTA	TTTAGTCCCT	ATGGAGTTCT	TGACCTTGTT	TTGAAGGGCG	GTAATCTGTT	1740
GCAAAATATT	AAAGATGTTC	ATGGAGATAC	AGATGACATA	AAACACATTA	AGAACTGTT	1800
GGATGAGGAA	GACGCAGTAG	CAGTTGTTCT	TGGTGGCAAG	GACAATACCA	CAATTGATAA	1860
ATTACTACAA	CACGAAAAAG	AACAAGCAGA	ACAATGCAAA	CAAAAGCAGG	AAGAATGCGA	1920
GAAAAAAGCA	CAACAAGAAA	GTCGTGGTCG	CTCCGCCGAA	ACCCGCGAAG	ACGAAAGGAC	1980
ACAACAACCT	GCTGATAGTG	CCGGCGAAGT	CGAAGAAGAA	GAAGACGACG	ACGACTACGA	2040
CGAAGACGAC	GAAGATGACG	ACGTAGTCCA	GGACGTAGAT	GTAAGTGAAA	TAAGAGGTCC	2100
G						2101

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 700 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: *Plasmodium falciparum*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Glu Gln Gly Asp Asn Lys Val Gly Ala Cys Ala Pro Tyr Arg Arg Leu
 1 5 10 15
 His Leu Cys Asp Tyr Asn Leu Glu Ser Ile Asp Thr Thr Ser Thr Thr
 20 25 30
 His Lys Leu Leu Leu Glu Val Cys Met Ala Ala Lys Tyr Glu Gly Asn
 35 40 45
 Ser Ile Asn Thr His Tyr Thr Gln His Gln Arg Thr Asn Glu Asp Ser
 50 55 60
 Ala Ser Gln Leu Cys Thr Val Leu Ala Arg Ser Phe Ala Asp Ile Gly
 65 70 75 80
 Asp Ile Val Arg Gly Lys Asp Leu Tyr Leu Gly Tyr Asp Asn Lys Glu
 85 90 95
 Lys Glu Gln Arg Lys Lys Leu Glu Gln Lys Leu Lys Asp Ile Phe Lys
 100 105 110
 Lys Ile His Lys Asp Val Met Lys Thr Asn Gly Ala Gln Glu Arg Tyr
 115 120 125
 Ile Asp Asp Ala Lys Gly Gly Asp Phe Phe Gln Leu Arg Glu Asp Trp
 130 135 140
 Trp Thr Ser Asn Arg Glu Thr Val Trp Lys Ala Leu Ile Cys His Ala
 145 150 155 160
 Pro Lys Glu Ala Asn Tyr Phe Ile Lys Thr Ala Cys Asn Val Gly Lys
 165 170 175
 Gly Thr Asn Gly Gln Cys His Cys Ile Gly Gly Asp Val Pro Thr Tyr
 180 185 190
 Phe Asp Tyr Val Pro Gln Tyr Leu Arg Trp Phe Glu Glu Trp Ala Glu
 195 200 205
 Asp Phe Cys Arg Lys Lys Lys Lys Lys Leu Glu Asn Leu Gln Lys Gln
 210 215 220
 Cys Arg Asp Tyr Glu Gln Asn Leu Tyr Cys Ser Gly Asn Gly Tyr Asp
 225 230 235 240
 Cys Thr Lys Thr Ile Tyr Lys Lys Gly Lys Leu Val Ile Gly Glu His
 245 250 255

59

Cys Thr Asn Cys Ser Val Trp Cys Arg Met Tyr Glu Thr Trp Ile Asp
 260 265 270
 Asn Gln Lys Lys Glu Phe Leu Lys Gln Lys Arg Lys Tyr Glu Thr Glu
 275 280 285
 Ile Ser Gly Gly Gly Ser Gly Lys Ser Pro Lys Arg Thr Lys Arg Ala
 290 295 300
 Ala Arg Ser Ser Ser Ser Ser Asp Asp Asn Gly Tyr Glu Ser Lys Phe
 305 310 315 320
 Tyr Lys Lys Leu Lys Glu Val Gly Tyr Gln Asp Val Asp Lys Phe Leu
 325 330 335
 Lys Ile Leu Asn Lys Glu Gly Ile Cys Gln Lys Gln Pro Gln Val Gly
 340 345 350
 Asn Glu Lys Ala Asp Asn Val Asp Phe Thr Asn Glu Lys Tyr Val Lys
 355 360 365
 Thr Phe Ser Arg Thr Glu Ile Cys Glu Pro Cys Pro Trp Cys Gly Leu
 370 375 380
 Glu Lys Gly Gly Pro Pro Trp Lys Val Lys Gly Asp Lys Thr Cys Gly
 385 390 395 400
 Ser Ala Lys Thr Lys Thr Tyr Asp Pro Lys Asn Ile Thr Asp Ile Pro
 405 410 415
 Val Leu Tyr Pro Asp Lys Ser Gln Gln Asn Ile Leu Lys Lys Tyr Lys
 420 425 430
 Asn Phe Cys Glu Lys Gly Ala Pro Gly Gly Gly Gln Ile Lys Lys Trp
 435 440 445
 Gln Cys Tyr Tyr Asp Glu His Arg Pro Ser Ser Lys Asn Asn Asn Asn
 450 455 460
 Cys Val Glu Gly Thr Trp Asp Lys Phe Thr Gln Gly Lys Gln Thr Val
 465 470 475 480
 Lys Ser Tyr Asn Val Phe Phe Trp Asp Trp Val His Asp Met Leu His
 485 490 495
 Asp Ser Val Glu Trp Lys Thr Glu Leu Ser Lys Cys Ile Asn Asn Asn
 500 505 510
 Thr Asn Gly Asn Thr Cys Arg Asn Asn Asn Lys Cys Lys Thr Asp Cys
 515 520 525
 Gly Cys Phe Gln Lys Trp Val Glu Lys Lys Gln Gln Glu Trp Met Ala
 530 535 540
 Ile Lys Asp His Phe Gly Lys Gln Thr Asp Ile Val Gln Gln Lys Gly
 545 550 555 560
 Leu Ile Val Phe Ser Pro Tyr Gly Val Leu Asp Leu Val Leu Lys Gly
 565 570 575
 Gly Asn Leu Leu Gln Asn Ile Lys Asp Val His Gly Asp Thr Asp Asp
 580 585 590
 Ile Lys His Ile Lys Lys Leu Leu Asp Glu Glu Asp Ala Val Ala Val
 595 600 605

60

Val Leu Gly Gly Lys Asp Asn Thr Thr Ile Asp Lys Leu Leu Gln His
610 615 620

Glu Lys Glu Gln Ala Glu Gln Cys Lys Gln Lys Gln Glu Glu Cys Glu
625 630 635 640

Lys Lys Ala Gln Gln Glu Ser Arg Gly Arg Ser Ala Glu Thr Arg Glu
645 650 655

Asp Glu Arg Thr Gln Gln Pro Ala Asp Ser Ala Gly Glu Val Glu Glu
660 665 670

Glu Glu Asp Asp Asp Asp Tyr Asp Glu Asp Asp Glu Asp Asp Asp Val
675 680 685

Val Gln Asp Val Asp Val Ser Glu Ile Arg Gly Pro
690 695 700

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 8220 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Plasmodium falciparum*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAAAATGGGG	CCCAAGGAGG	CTGCAGGTGG	GGATGATATT	GAGGATGAAA	GTGCCAAACA	60
TATGTTTGAT	AGGATAGGAA	AAGATGTGTA	CGATAAGTA	AAAGAGGAAG	CTAAAGAACG	120
TGGTAAAGGC	TTGCAAGGAC	GTTTGTCAGA	AGCAAAATTT	GAGAAAAATG	AAAGCGATCC	180
ACAAACACCA	GAAGATCCAT	GCGATCTTGA	TCATAAATAT	CATACAAATG	TAACACTAA	240
TGTAATTAAT	CCGTGCGCTG	ATAGATCTGA	CGTGCGTTTT	TCCGATGAAT	ATGGAGGTCA	300
ATGTACACAT	AATAGAATAA	AAGATAGTCA	ACAGGGTGAT	AATAAAGGTG	CATGTGCTCC	360
ATATAGGCGA	TTGCATGTAT	GCGATCAAAA	TTTAGAACAG	ATAGAGCCTA	TAAAAATAAC	420
AAATACTCAT	AATTTATTGG	TAGATGTGTG	TATGGCAGCA	AAATTTGAAG	GACAATCAAT	480
AACACAAGAT	TATCCAAAAT	ATCAAGCAAC	ATATGGTGAT	TCTCCTTCTC	AAATATGTAC	540
TATGCTGGCA	CGAAGTTTTG	CGGACATAGG	GGACATTGTC	AGAGGAAGAG	ATTTGTATTT	600
AGGTAATCCA	CAAGAAATAA	AACAAAGACA	ACAATTAGAA	AATAATTTGA	AAACAATTTT	660
CGGGAAAATA	TATGAAAAT	TGAATGGCGC	AGAAGCACGC	TACGGAAATG	ATCCGGAATT	720
TTTTAAATTA	CGAGAAGATT	GGTGGACTGC	TAATCGAGAA	ACAGTATGGA	AAGCCATCAC	780
ATGTAACGCT	TGGGGTAATA	CATATTTTCA	TGCAACGTGC	AATAGAGGAG	AACGAACTAA	840
AGGTTACTGC	CGGTGTAACG	ACGACCAAGT	TCCCACATAT	TTTGATTATG	TGCCGCAGTA	900
TCTTCGCTGG	TTGAGGAAT	GGGCAGAAGA	TTTTTGTAGG	AAAAAAAATA	AAAAATAAA	960
AGATGTTAAA	AGAAATTGTC	GTGGAAAAGA	TAAAGAGGAT	AAGGATCGAT	ATTGTAGCCG	1020
TAATGGCTAC	GATTGCGAAA	AAACTAAACG	AGCGATTGGT	AAGTTGCGTT	ATGGTAAGCA	1080
ATGCATTAGC	TGTTTGATG	CATGTAATCC	TTACGTTGAT	TGGATAAATA	ACCAAAAAGA	1140
ACAATTTGAC	AAACAGAAAA	AAAAATATGA	TGAAGAAATA	AAAAAATATG	AAAATGGAGC	1200
ATCAGGTGGT	AGTAGGCAAA	AACGGGATGC	AGGTGGTACA	ACTACTACTA	ATTATGATGG	1260
ATATGAAAAA	AAATTTTATG	ACGAACTTAA	TAAAAGTGAA	TATAGAACCG	TTGATAAATT	1320
TTTGGA AAAA	TTAAGTAATG	AAGAAATATG	CACAAAAGTT	AAAGACGAAG	AAGGAGGAAC	1380
AATTGATTTT	AAAAACGTTA	ATAGTGATAG	TACTAGTGGT	GCTAGTGGCA	CTAATGTTGA	1440
AAGTCAAGGA	ACATTTTATC	GTTCAAAATA	TTGCCAACCC	TGCCCTTATT	GTGGAGTGAA	1500

AAAGGTAAAT	AATGGTGGTA	GTAGTAATGA	ATGGGAAGAG	AAAAATAATG	GCAAGTGCAA	1560
GAGTGGAAAA	CTTTATGAGC	CTAAACCCGA	CAAAGAAGGT	ACTACTATTA	CAATCCTTAA	1620
AAGTGGTAAA	GGACATGATG	ATATTGAAGA	AAAATTAAAC	AAATTTTGTG	ATGAAAAAAA	1680
TGGTGATACA	ATAAATAGTG	GTGGTAGTGG	TACGGGTGGT	AGTGGTGGTG	GTAACAGTGG	1740
TAGACAGGAA	TTGTATGAAG	AATGGAAATG	TTATAAAGGT	GAAGATGTAG	TGAAAGTTGG	1800
ACACGATGAG	GATGACGAGG	AGGATTATGA	AAATGTAAAA	AATGCAGGCG	GATTATGTAT	1860
ATTAAAAAAC	CAAAAAAGA	ATAAGAAGA	AGGTGGAAAT	ACGTCTGAAA	AGGAGCCTGA	1920
TGAAATCCAA	AAGACATTCA	ATCCTTTTTT	TTACTATTGG	GTTGCACATA	TGTTAAAAGA	1980
TTCCATACAT	TGAAAAAAA	AACCTCAGAG	ATGTTTACAA	AATGGTAACA	GAATAAAATG	2040
TGGAAACAAT	AAATGTAATA	ATGATTGTGA	ATGTTTTAAA	AGATGGATTA	CACAAAAAAA	2100
AGACGAATGG	GGGAAAATAG	TACAACATTT	TAAACGCAA	AATATTAAAG	GTAGAGGAGG	2160
TAGTGACAAT	ACGGCAGAAT	TAATCCCATT	TGATCACGAT	TATGTTCTTC	AATACAATTT	2220
GCAAGAAGAA	TTTTTGAAAG	GCGATTCCGA	AGACGCTTCC	GAAGAAAAAT	CCGAAAATAG	2280
TCTGGATGCA	GAGGAGGCAG	AGGAACTAAA	ACACCTTCGC	GAAATCATTG	AAAGTGAAGA	2340
CAATAATCAA	GAAGCATCTG	TTGGTGGTGG	CGTCACTGAA	CAAAAAATA	TAATGGATAA	2400
ATTGCTCAAC	TACGAAAAAG	ACGAAGCCGA	TTTATGCCTA	GAAATTCACG	AAGATGAGGA	2460
AGAGGAAAAA	GAAAAAGGAG	ACGGAACGA	ATGTATCGAA	GAGGGCGAAA	ATTTTCGTTA	2520
TAATCCATGT	AGTGGCGAAA	GTGGTAACAA	ACGATACCCC	GTTCTTGCGA	ACAAAGTAGC	2580
GTATCAAATG	CATCACAAGG	CAAAGACACA	ATTGGCTAGT	CGTGCTGGTA	GAAGTGCGTT	2640
GAGAGGTGAT	ATATCCTTAG	CGCAATTTAA	AAATGGTCGT	AACGGAAGTA	CATTGAAAGG	2700
ACAAATTTGC	AAAATTAACG	AAACTATTTC	CAATGATAGT	CGTGGTAATA	GTGGTGGACC	2760
ATGTACAGGC	AAAGATGGAG	ATCACGGAGG	TGTGCGCATG	AGAATAGGAA	CGGAATGGTC	2820
AAATATTGAA	GGAAAAAAC	AAACGTCATA	CAAAAACGTC	TTTTTACCTC	CCCGACGAGA	2880
ACACATGTGT	ACATCCAATT	TAGAAAATTT	AGATGTTGGT	AGTGTCACCTA	AAAATGATAA	2940
GGCTAGCCAC	TCATTATTGG	GAGATGTTCA	GCTCGCAGCA	AAAACCTGATG	CAGCTGAGAT	3000
AATAAACGCG	TATAAAGATC	AAAATAATAT	ACAACTAACT	GATCCAATAC	AACAAAAAGA	3060
CCAGGAGGCT	ATGTGTCGAG	CTGTACGTTA	TAGTTTTGCC	GATTTAGGAG	ACATTATTCCG	3120
AGGAAGAGAT	ATGTGGGATG	AGGATAAGAG	CTCAACAGAC	ATGGAAACAC	GTTTGATAAC	3180
CGTATTTAAA	AACATTAAAG	AAAAACATGA	TGGAATCAAA	GACAACCCTA	AATATACCGG	3240
TGATGAAAGC	AAAAAGCCCG	CATATAAAAA	ATTACGAGCA	GATTGGTGGG	AAGCAAATAG	3300
ACATCAAGTG	TGGAGAGCCA	TGAAATGCGC	AACAAAAGGC	ATCATATGTC	CTGGTATGCC	3360
AGTTGACGAT	TATATCCCCC	AACGTTTACG	CTGGATGACT	GAATGGGCTG	AATGGTATTG	3420
TAAAGCGCAA	TCACAGGAGT	ATGACAAGTT	AAAAAAAATC	TGTGCAGATT	GTATGAGTAA	3480
GGGTGATGGA	AAATGTACGC	AAGGTGATGT	CGATTGTGGA	AAGTGCAAAG	CAGCATGTGA	3540

TAAATATAAA	GAGGAAATAG	AAAAATGGAA	TGAACAATGG	AGAAAAATAT	CAGATAAATA	3600
CAATCTATTA	TACCTACAAG	CAAAAACACTAC	TTCTACTAAT	CCTGGCCGTA	CTGTTCTTGG	3660
TGATGACGAT	CCCGACTATC	AACAAATGGT	AGATTTTTTG	ACCCAATAC	ACAAAGCAAG	3720
TATTGCCGCA	CGTGTTCTTG	TTAAACGTGC	TGCTGGTAGT	CCCCTGAGA	TCGCCGCCGC	3780
CGCCCCGATC	ACCCCTACA	GTACTGCTGC	CGGATATATA	CACCAGGAAA	TAGGATATGG	3840
GGGGTGCCAG	GAACAAACAC	AATTTTGTGA	AAAAAACAT	GGTGCAACAT	CAACTAGTAC	3900
CACGAAAGAA	AACAAAGAAT	ACACCTTTAA	ACAACCTCCG	CCGGAGTATG	CTACAGCGTG	3960
TGATTGCATA	AATAGGTCGC	AAACAGAGGA	GGCGAAGAAA	AAGGAAGAAA	ATGTAGAGAG	4020
TGCCTGCAAA	ATAGTGGAGA	AAATACTTGA	GGGTAAGAAT	GGAAGGACTA	CAGTAGGTGA	4080
ATGTAATCCA	AAAGAGAGTT	ATCCTGATTG	GGATTGCAAA	AACAATATTG	ACATTAGTCA	4140
TGATGGTGCT	TGTATGCCTC	CAAGGAGACA	AAAACATATG	TTATATTATA	TAGCACATGA	4200
GAGTCAAACA	GAAAATATAA	AAACAGACGA	TAATTTGAAA	GATGCTTTTA	TTAAACTGC	4260
AGCAGCAGAA	ACTTTTCTTT	CATGGCAATA	TTATAAGAGT	AAGAATGATA	GTGAAGCTAA	4320
AATATTAGAT	AGAGGCCTTA	TTCCATCCCA	ATTTTAAAGA	TCCATGATGT	ACACGTTTGG	4380
AGATTATAGA	GATATATGTT	TGAACACAGA	TATATCTAAA	AAACAAAATG	ATGTAGCTAA	4440
GGCAAAAGAT	AAAATAGGTA	AATTTTTCTC	AAAAGATGGC	AGCAAATCTC	CTAGTGGCTT	4500
ATCACGCCAA	GAATGGTGGA	AAACAAATGG	TCCAGAGATT	TGGAAAGGAA	TGTTATGTGC	4560
CTTAACAAAA	TACGTCACAG	ATACCGATAA	CAAAAGAAAA	ATCAAAAACG	ACTACTCATA	4620
CGATAAAGTC	AACCAATCCC	AAAATGGCAA	CCCTTCCCTT	GAAGAGTTTG	CTGCTAAACC	4680
TCAATTTCTA	CGTTGGATGA	TCGAATGGGG	AGAAGAGTTT	TGTGCTGAAC	GTCAGAAGAA	4740
GGAAAATATC	ATAAAAGATG	CATGTAATGA	AATAAATTCT	ACACAACAGT	GTAATGATGC	4800
GAAACATCGT	TGTAATCAAG	CATGTAGAGC	ATATCAAGAA	TATGTTGAAA	ATAAAAAAAA	4860
AGAATTTTCG	GGACAAACAA	ATAACTTTGT	TCTAAAGGCA	AATGTTTCAGC	CCCAAGATCC	4920
AGAATATAAA	GGATATGAAT	ATAAAGACGG	CGTACAACCG	ATACAGGGGA	ATGAGTATTT	4980
ACTGCAAAAA	TGTGATAATA	ATAAATGTTC	TTGCATGGAT	GGAAATGTAC	TTTCCGTCTC	5040
TCCAAAAGAA	AAACCTTTTG	GAAAATATGC	CCATAAATAT	CCTGAGAAAT	GTGATTGTTA	5100
TCAAGGAAAA	CATGTACCTA	GCATACCACC	TCCCCCCCCA	CCTGTACAAC	CACAACCGGA	5160
AGCACCAACA	GTAACAGTAG	ACGTTTGCAG	CATAGTAAAA	ACACTATTTA	AAGACACAAA	5220
CAATTTTTC	GACGCTTGTG	GTCTAAAATA	CGGCAAAACC	GCACCATCCA	GTTGGAAATG	5280
TATACCAAGT	GACACAAAAA	GTGGTGCTGG	TGCCACCACC	GGCAAAAGTG	GTAGTGATAG	5340
TGGTAGTATT	TGTATCCAC	CCAGGAGGCG	ACGATTATAT	GTGGGGAAAC	TACAGGAGTG	5400
GGCTACCGCG	CTCCACAAAG	GTGAGGGCGC	CGCGCCGTCC	CACTCACGCG	CCGACGACTT	5460
GCGCAATGCG	TTCATCCAAT	CTGCTGCAAT	AGAGACTTTT	TTCTTATGGG	ATAGATATAA	5520
AGAAGAGAAA	AAACCACAGG	GTGATGGGTC	ACAACAAGCA	CTATCACAAC	TAACCAGTAC	5580

ATACAGTGAT	GACGAGGAGG	ACCCCCCGA	CAAACGTGTA	CAAAATGGTA	AGATACCCCC	5640
CGATTTTTTG	AGATTAATGT	TCTATACATT	AGGAGATTAT	AGGGATATTT	TAGTACACGG	5700
TGGTAACACA	AGTGACAGTG	GTAACACAAA	TGGTAGTAAC	AACAACAATA	TTGTGCTTGA	5760
AGCGAGTGGT	AACAAGGAGG	ACATGCAAAA	AATACAAGAG	AAAATAGAAC	AAATTCTCCC	5820
AAAAAATGGT	GGCACACCTC	TTGTCCCAAA	ATCTAGTGCC	CAAACACCTG	ATAAATGGTG	5880
GAATGAACAC	GCCGAATCTA	TCTGGAAAGG	TATGATATGT	GCATTGACAT	ATACAGAAAA	5940
GAACCCTGAC	ACCAGTGCAA	GAGGCGACGA	AAACAAAATA	GAAAAGGATG	ATGAAGTGTA	6000
CGAGAAATTT	TTTGGCAGCA	CAGCCGACAA	ACATGGCACA	GCCTCAACCC	CAACCGGCAC	6060
ATACAAAACC	CAATACGACT	ACGAAAAAGT	CAAACCTGAG	GATACAAGTG	GTGCCAAAAAC	6120
CCCCTCAGCC	TCTAGTGATA	CACCCCTTCT	CTCCGATTTT	GTGTTACGCC	CCCCCTACTT	6180
CCGTTACCTT	GAAGAATGGG	GTCAAAATTT	TTGTAAAAAA	AGAAAGCATA	AATTGGCACA	6240
AATAAAACAT	GAGTGTAAG	TAGAAGAAAA	TGGTGGTGGT	AGTCGTCGTG	GTGGTATAAC	6300
AAGACAATAT	AGTGGGGATG	GCGAAGCGTG	TAATGAGATG	CTTCCAAAAA	ACGATGGAAC	6360
TGTTCCGGAT	TTAGAAAAGC	CGAGTTGTGC	CAAACCTTGT	AGTTCTTATA	GAAAATGGAT	6420
AGAAAGCAAG	GGAAAAGAGT	TTGAGAAACA	AGAAAAGGCA	TATGAACAAC	AAAAAGACAA	6480
ATGTGTAAAT	GGAAGTAATA	AGCATGATAA	TGGATTTTGT	GAAACACTAA	CAACGTCCTC	6540
TAAAGCTAAA	GACTTTTTTA	AAACGTTAGG	ACCATGTAAA	CCTAATAATG	TAGAGGGTAA	6600
AACAATTTTT	GATGATGATA	AAACCTTTAA	ACATACAAAA	GATTGTGATC	CATGTCTTAA	6660
ATTTAGTGTT	AATTGTAAAA	AAGATGAATG	TGATAATTCT	AAAGGAACCG	ATTGCCGAAA	6720
TAAAAATAGT	ATTGATGCAA	CAGATATTGA	AAATGGAGTG	GATTCTACTG	TACTAGAAAT	6780
GCGTGTCAAG	GCTGATAGTA	AAAGTGGATT	TAATGGTGAT	GGTTTAGAGA	ATGCTTGTAG	6840
AGGTGCTGGT	ATCTTTGAAG	GTATTAGAAA	AGATGAATGG	AAATGTCGTA	ATGTATGTGG	6900
TTATGTTGTA	TGTAAACCGG	AAAACGTTAA	TGGGGAAGCA	AAGGGAAAAC	ACATTATACA	6960
AATTAGAGCA	CTGGTTAAAC	GTTGGGTAGA	ATATTTTTTT	GAAGATTATA	ATAAAATAAA	7020
ACATAAAATT	TCACATCGCA	TAAAAAATGG	TGAAATATCT	CCATGTATAA	AAAATTGTGT	7080
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TCAATATAAA	AATGACAATT	CAGATGATGA	CAATGTGAGA	AGTTTTTTGG	AGACCTTGAT	7200
ACCTCAAATT	ACTGATGCAA	ACGCTAAAAA	TAAGGTTATA	AAATTAAGTA	AGTTCGGTAA	7260
TTCTTGTTGA	TGTAGTGCCA	GTGCGAACGA	ACAAAACAAA	AATGGTGAAT	ACAAGGACGC	7320
TATAGATTGT	ATGCTTAAAA	AGCTTAAAGA	TAAAATTGGC	GAGTGCGAAA	AGAAACACCA	7380
TCAAAC TAGT	GATACCGAGT	GTTCCGACAC	ACCACAACCG	CAAACCCCTG	AAGACGAAAC	7440
TTTGGATGAT	GATATAGAAA	CAGAGGAGGC	GAAGAAGAAC	ATGATGCCGA	AAATTTGTGA	7500
AAATGTGTTA	AAAACAGCAC	AACAAGAGGA	TGAAGGCGGT	TGTGTCCCAG	CAGAAAATAG	7560
TGAAGAACCG	GCAGCAACAG	ATAGTGGTAA	GGAAACCCCC	GAACAAACCC	CCGTTCTCAA	7620

ACCCGAAGAA	GAAGCAGTAC	CGGAACCACC	ACCTCCACCC	CCACAGGAAA	AAGCCCCGGC	7680
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TCTAACCGCC	CTGGTGACCT	CCACCCTCGC	CTGGAGCGTT	GGCATCGGTT	TTGCTACATT	7800
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TTGTGATTAT	GTTTGGATAT	ATATATATAT	ATATATATGT	TTATGTATAT	GTGTTTTTGG	7980
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TATATGGATG	TACTTGTATG	TGTTTTATAT	ATATATTTTA	TATATATGTA	TTTATATTAA	8100
AAAAGAAATA	TAAAAACAAA	TTTATTAAAA	TGAAAAAAG	AAAAATGAAA	TATAAAAAAA	8160
AATTTATTAA	AATAAAAAAA	AAAAAAAAAA	AAAAGGAGAA	AAATTTTTTA	AAAAATAATA	8220

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2710 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

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Asn Val Met Val Glu Leu Ala Lys Met Gly Pro Lys Glu Ala Ala Gly
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          20           25           30
Gly Lys Asp Val Tyr Asp Lys Val Lys Glu Glu Ala Lys Glu Arg Gly
          35           40           45
Lys Gly Leu Gln Gly Arg Leu Ser Glu Ala Lys Phe Glu Lys Asn Glu
          50           55           60
Ser Asp Pro Gln Thr Pro Glu Asp Pro Cys Asp Leu Asp His Lys Tyr
          65           70           75           80
His Thr Asn Val Thr Thr Asn Val Ile Asn Pro Cys Ala Asp Arg Ser
          85           90           95
Asp Val Arg Phe Ser Asp Glu Tyr Gly Gly Gln Cys Thr His Asn Arg
          100          105          110
Ile Lys Asp Ser Gln Gln Gly Asp Asn Lys Gly Ala Cys Ala Pro Tyr
          115          120          125
Arg Arg Leu His Val Cys Asp Gln Asn Leu Glu Gln Ile Glu Pro Ile
          130          135          140
Lys Ile Thr Asn Thr His Asn Leu Leu Val Asp Val Cys Met Ala Ala
          145          150          155          160
Lys Phe Glu Gly Gln Ser Ile Thr Gln Asp Tyr Pro Lys Tyr Gln Ala
          165          170          175
Thr Tyr Gly Asp Ser Pro Ser Gln Ile Cys Thr Met Leu Ala Arg Ser
          180          185          190
Phe Ala Asp Ile Gly Asp Ile Val Arg Gly Arg Asp Leu Tyr Leu Gly
          195          200          205
Asn Pro Gln Glu Ile Lys Gln Arg Gln Gln Leu Glu Asn Asn Leu Lys
          210          215          220
Thr Ile Phe Gly Lys Ile Tyr Glu Lys Leu Asn Gly Ala Glu Ala Arg
          225          230          235          240
Tyr Gly Asn Asp Pro Glu Phe Phe Lys Leu Arg Glu Asp Trp Trp Thr
          245          250          255

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Ala Asn Arg Glu Thr Val Trp Lys Ala Il Thr Cys Asn Ala Trp Gly
 260 265 270
 Asn Thr Tyr Phe His Ala Thr Cys Asn Arg Gly Glu Arg Thr Lys Gly
 275 280 285
 Tyr Cys Arg Cys Asn Asp Asp Gln Val Pro Thr Tyr Phe Asp Tyr Val
 290 295 300
 Pro Gln Tyr Leu Arg Trp Phe Glu Glu Trp Ala Glu Asp Phe Cys Arg
 305 310 315 320
 Lys Lys Asn Lys Lys Ile Lys Asp Val Lys Arg Asn Cys Arg Gly Lys
 325 330 335
 Asp Lys Glu Asp Lys Asp Arg Tyr Cys Ser Arg Asn Gly Tyr Asp Cys
 340 345 350
 Glu Lys Thr Lys Arg Ala Ile Gly Lys Leu Arg Tyr Gly Lys Gln Cys
 355 360 365
 Ile Ser Cys Leu Tyr Ala Cys Asn Pro Tyr Val Asp Trp Ile Asn Asn
 370 375 380
 Gln Lys Glu Gln Phe Asp Lys Gln Lys Lys Tyr Asp Glu Glu Ile
 385 390 395 400
 Lys Lys Tyr Glu Asn Gly Ala Ser Gly Gly Ser Arg Gln Lys Arg Asp
 405 410 415
 Ala Gly Gly Thr Thr Thr Thr Asn Tyr Asp Gly Tyr Glu Lys Lys Phe
 420 425 430
 Tyr Asp Glu Leu Asn Lys Ser Glu Tyr Arg Thr Val Asp Lys Phe Leu
 435 440 445

68

Glu Lys Leu Ser Asn Glu Glu Ile Cys Thr Lys Val Lys Asp Glu Glu
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Gly Gly Thr Ile Asp Phe Lys Asn Val Asn Ser Asp Ser Thr Ser Gly
465 470 475 480
Ala Ser Gly Thr Asn Val Glu Ser Gln Gly Thr Phe Tyr Arg Ser Lys
485 490 495
Tyr Cys Gln Pro Cys Pro Tyr Cys Gly Val Lys Lys Val Asn Asn Gly
500 505 510
Gly Ser Ser Asn Glu Trp Glu Glu Lys Asn Asn Gly Lys Cys Lys Ser
515 520 525
Gly Lys Leu Tyr Glu Pro Lys Pro Asp Lys Glu Gly Thr Thr Ile Thr
530 535 540
Ile Leu Lys Ser Gly Lys Gly His Asp Asp Ile Glu Glu Lys Leu Asn
545 550 555 560
Lys Phe Cys Asp Glu Lys Asn Gly Asp Thr Ile Asn Ser Gly Gly Ser
565 570 575
Gly Thr Gly Gly Ser Gly Gly Gly Asn Ser Gly Arg Gln Glu Leu Tyr
580 585 590
Glu Glu Trp Lys Cys Tyr Lys Gly Glu Asp Val Val Lys Val Gly His
595 600 605

Asp Glu Asp Asp Glu Glu Asp Tyr Glu Asn Val Lys Asn Ala Gly Gly
 610 615 620
 Leu Cys Ile Leu Lys Asn Gln Lys Lys Asn Lys Glu Glu ly Gly Asn
 625 630 635 640
 Thr Ser Glu Lys Glu Pro Asp Glu Ile Gln Lys Thr Phe Asn Pro Phe
 645 650 655
 Phe Tyr Tyr Trp Val Ala His Met Leu Lys Asp Ser Ile His Trp Lys
 660 665 670
 Lys Lys Leu Gln Arg Cys Leu Gln Asn Gly Asn Arg Ile Lys Cys Gly
 675 680 685
 Asn Asn Lys Cys Asn Asn Asp Cys Glu Cys Phe Lys Arg Trp Ile Thr
 690 695 700
 Gln Lys Lys Asp Glu Trp Gly Lys Ile Val Gln His Phe Lys Thr Gln
 705 710 715 720
 Asn Ile Lys Gly Arg Gly Gly Ser Asp Asn Thr Ala Glu Leu Ile Pro
 725 730 735
 Phe Asp His Asp Tyr Val Leu Gln Tyr Asn Leu Gln Glu Glu Phe Leu
 740 745 750
 Lys Gly Asp Ser Glu Asp Ala Ser Glu Glu Lys Ser Glu Asn Ser Leu
 755 760 765
 Asp Ala Glu Glu Ala Glu Glu Leu Lys His Leu Arg Glu Ile Ile Glu
 770 775 780
 Ser Glu Asp Asn Asn Gln Glu Ala Ser Val Gly Gly Gly Val Thr Glu
 785 790 795 800
 Gln Lys Asn Ile Met Asp Lys Leu Leu Asn Tyr Glu Lys Asp Glu Ala
 805 810 815
 Asp Leu Cys Leu Glu Ile His Glu Asp Glu Glu Glu Glu Lys Glu Lys
 820 825 830
 Gly Asp Gly Asn Glu Cys Ile Glu Glu Gly Glu Asn Phe Arg Tyr Asn
 835 840 845
 Pro Cys Ser Gly Glu Ser Gly Asn Lys Arg Tyr Pro Val Leu Ala Asn
 850 855 860
 Lys Val Ala Tyr Gln Met His His Lys Ala Lys Thr Gln Leu Ala Ser
 865 870 875 880
 Arg Ala Gly Arg Ser Ala Leu Arg Gly Asp Ile Ser Leu Ala Gln Phe
 885 890 895
 Lys Asn Gly Arg Asn Gly Ser Thr Leu Lys Gly Gln Ile Cys Lys Ile
 900 905 910
 Asn Glu Asn Tyr Ser Asn Asp Ser Arg Gly Asn Ser Gly Gly Pro Cys
 915 920 925
 Thr Gly Lys Asp Gly Asp His Gly Gly Val Arg Met Arg Ile Gly Thr
 930 935 940
 Glu Trp Ser Asn Ile Glu Gly Lys Lys Gln Thr Ser Tyr Lys Asn Val
 945 950 955 960

70

Phe Leu Pro Pro Arg Arg Glu His Met Cys Thr Ser Asn Leu Glu Asn
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 L u Asp Val Gly Ser Val Thr Lys Asn Asp Lys Ala Ser His S r L u
 980 985 990
 Leu Gly Asp Val Gln Leu Ala Ala Lys Thr Asp Ala Ala Glu Ile Ile
 995 1000 1005
 Lys Arg Tyr Lys Asp Gln Asn Asn Ile Gln Leu Thr Asp Pro Ile Gln
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 1045 1050 1055
 Ser Ser Thr Asp Met Glu Thr Arg Leu Ile Thr Val Phe Lys Asn Ile
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 1075 1080 1085
 Glu Ser Lys Lys Pro Ala Tyr Lys Lys Leu Arg Ala Asp Trp Trp Glu
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 Ile Ile Cys Pro Gly Met Pro Val Asp Asp Tyr Ile Pro Gln Arg Leu
 1125 1130 1135
 Arg Trp Met Thr Glu Trp Ala Glu Trp Tyr Cys Lys Ala Gln Ser Gln
 1140 1145 1150
 Glu Tyr Asp Lys Leu Lys Lys Ile Cys Ala Asp Cys Met Ser Lys Gly
 1155 1160 1165
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 1170 1175 1180
 Ala Cys Asp Lys Tyr Lys Glu Glu Ile Glu Lys Trp Asn Glu Gln Trp
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 Thr Ser Thr Asn Pro Gly Arg Thr Val Leu Gly Asp Asp Asp Pro Asp
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 Tyr Gln Gln Met Val Asp Phe Leu Thr Pro Ile His Lys Ala Ser Ile
 1235 1240 1245
 Ala Ala Arg Val Leu Val Lys Arg Ala Ala Gly Ser Pro Thr Glu Ile
 1250 1255 1260
 Ala Ala Ala Ala Pro Ile Thr Pro Tyr Ser Thr Ala Ala Gly Tyr Ile
 1265 1270 1275 1280
 His Gln Glu Ile Gly Tyr Gly Gly Cys Gln Glu Gln Thr Gln Phe Cys
 1285 1290 1295
 Glu Lys Lys His Gly Ala Thr Ser Thr Ser Thr Thr Lys Glu Asn Lys
 1300 1305 1310

Glu Tyr Thr Phe Lys Gln Pro Pro Pro Glu Tyr Ala Thr Ala Cys Asp
 1315 1320 1325
 Cys Ile Asn Arg Ser Gln Thr Glu Glu Pro Lys Lys Lys Glu Glu Asn
 1330 1335 1340
 Val Glu Ser Ala Cys Lys Ile Val Glu Lys Ile Leu Glu Gly Lys Asn
 1345 1350 1355 1360
 Gly Arg Thr Thr Val Gly Glu Cys Asn Pro Lys Glu Ser Tyr Pro Asp
 1365 1370 1375
 Trp Asp Cys Lys Asn Asn Ile Asp Ile Ser His Asp Gly Ala Cys Met
 1380 1385 1390
 Pro Pro Arg Arg Gln Lys Leu Cys Leu Tyr Tyr Ile Ala His Glu Ser
 1395 1400 1405
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 Lys Asp Lys Ile Gly Lys Phe Phe Ser Lys Asp Gly Ser Lys Ser Pro
 1490 1495 1500
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 1505 1510 1515 1520
 Trp Lys Gly Met Leu Cys Ala Leu Thr Lys Tyr Val Thr Asp Thr Asp
 1525 1530 1535
 Asn Lys Arg Lys Ile Lys Asn Asp Tyr Ser Tyr Asp Lys Val Asn Gln
 1540 1545 1550
 Ser Gln Asn Gly Asn Pro Ser Leu Glu Glu Phe Ala Ala Lys Pro Gln
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 Phe Leu Arg Trp Met Ile Glu Trp Gly Glu Glu Phe Cys Ala Glu Arg
 1570 1575 1580
 Gln Lys Lys Glu Asn Ile Ile Lys Asp Ala Cys Asn Glu Ile Asn Ser
 1585 1590 1595 1600
 Thr Gln Gln Cys Asn Asp Ala Lys His Arg Cys Asn Gln Ala Cys Arg
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 Ala Tyr Gln Glu Tyr Val Glu Asn Lys Lys Lys Glu Phe Ser Gly Gln
 1620 1625 1630
 Thr Asn Asn Phe Val Leu Lys Ala Asn Val Gln Pro Gln Asp Pro Glu
 1635 1640 1645
 Tyr Lys Gly Tyr Glu Tyr Lys Asp Gly Val Gln Pro Ile Gln Gly Asn
 1650 1655 1660

Glu Tyr Leu Leu Gln Lys Cys Asp Asn Asn Lys Cys Ser Cys Met Asp
 1665 1670 1675 1680
 Gly Asn Val Leu Ser Val Ser Pro Lys Glu Lys Pro Phe Gly Lys Tyr
 1685 1690 1695
 Ala His Lys Tyr Pro Glu Lys Cys Asp Cys Tyr Gln Gly Lys His Val
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 Pro Ser Ile Pro Pro Pro Pro Pro Val Gln Pro Gln Pro Glu Ala
 1715 1720 1725
 Pro Thr Val Thr Val Asp Val Cys Ser Ile Val Lys Thr Leu Phe Lys
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 1745 1750 1755 1760
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 1765 1770 1775
 Gly Ala Thr Thr Gly Lys Ser Gly Ser Asp Ser Gly Ser Ile Cys Ile
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 Phe Leu Trp Asp Arg Tyr Lys Glu Glu Lys Lys Pro Gln Gly Asp Gly
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 Ser Gln Gln Ala Leu Ser Gln Leu Thr Ser Thr Tyr Ser Asp Asp Glu
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 Glu Asp Pro Pro Asp Lys Leu Leu Gln Asn Gly Lys Ile Pro Pro Asp
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 Phe Leu Arg Leu Met Phe Tyr Thr Leu Gly Asp Tyr Arg Asp Ile Leu
 1890 1895 1900
 Val His Gly Gly Asn Thr Ser Asp Ser Gly Asn Thr Asn Gly Ser Asn
 1905 1910 1915 1920
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 1925 1930 1935
 Lys Ile Gln Glu Lys Ile Glu Gln Ile Leu Pro Lys Asn Gly Gly Thr
 1940 1945 1950
 Pro Leu Val Pro Lys Ser Ser Ala Gln Thr Pro Asp Lys Trp Trp Asn
 1955 1960 1965
 Glu His Ala Glu Ser Ile Trp Lys Gly Met Ile Cys Ala Leu Thr Tyr
 1970 1975 1980
 Thr Glu Lys Asn Pro Asp Thr Ser Ala Arg Gly Asp Glu Asn Lys Ile
 1985 1990 1995 2000
 Glu Lys Asp Asp Glu Val Tyr Glu Lys Phe Phe Gly Ser Thr Ala Asp
 2005 2010 2015

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Lys His Gly Thr Ala Ser Thr Pro Thr Gly Thr Tyr Lys Thr Gln Tyr
 2020 2025 2030
 Asp Tyr Glu Lys Val Lys Leu Glu Asp Thr Ser Gly Ala Lys Thr Pro
 2035 2040 2045
 Ser Ala Ser Ser Asp Thr Pro Leu Leu Ser Asp Phe Val Leu Arg Pro
 2050 2055 2060
 Pro Tyr Phe Arg Tyr Leu Glu Glu Trp Gly Gln Asn Phe Cys Lys Lys
 2065 2070 2075 2080
 Arg Lys His Lys Leu Ala Gln Ile Lys His Glu Cys Lys Val Glu Glu
 2085 2090 2095
 Asn Gly Gly Gly Ser Arg Arg Gly Gly Ile Thr Arg Gln Tyr Ser Gly
 2100 2105 2110
 Asp Gly Glu Ala Cys Asn Glu Met Leu Pro Lys Asn Asp Gly Thr Val
 2115 2120 2125
 Pro Asp Leu Glu Lys Pro Ser Cys Ala Lys Pro Cys Ser Ser Tyr Arg
 2130 2135 2140
 Lys Trp Ile Glu Ser Lys Gly Lys Glu Phe Glu Lys Gln Glu Lys Ala
 2145 2150 2155 2160
 Tyr Glu Gln Gln Lys Asp Lys Cys Val Asn Gly Ser Asn Lys His Asp
 2165 2170 2175
 Asn Gly Phe Cys Glu Thr Leu Thr Thr Ser Ser Lys Ala Lys Asp Phe
 2180 2185 2190
 Leu Lys Thr Leu Gly Pro Cys Lys Pro Asn Asn Val Glu Gly Lys Thr
 2195 2200 2205
 Ile Phe Asp Asp Asp Lys Thr Phe Lys His Thr Lys Asp Cys Asp Pro
 2210 2215 2220
 Cys Leu Lys Phe Ser Val Asn Cys Lys Lys Asp Glu Cys Asp Asn Ser
 2225 2230 2235 2240
 Lys Gly Thr Asp Cys Arg Asn Lys Asn Ser Ile Asp Ala Thr Asp Ile
 2245 2250 2255
 Glu Asn Gly Val Asp Ser Thr Val Leu Glu Met Arg Val Ser Ala Asp
 2260 2265 2270
 Ser Lys Ser Gly Phe Asn Gly Asp Gly Leu Glu Asn Ala Cys Arg Gly
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 Ala Gly Ile Phe Glu Gly Ile Arg Lys Asp Glu Trp Lys Cys Arg Asn
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 2305 2310 2315 2320
 Lys Gly Lys His Ile Ile Gln Ile Arg Ala Leu Val Lys Arg Trp Val
 2325 2330 2335
 Glu Tyr Phe Phe Glu Asp Tyr Asn Lys Ile Lys His Lys Ile Ser His
 2340 2345 2350
 Arg Ile Lys Asn Gly Glu Ile Ser Pro Cys Ile Lys Asn Cys Val Glu
 2355 2360 2365

Lys Trp Val Asp Gln Lys Arg Lys Glu Trp Lys Glu Ile Thr Glu Arg
 2370 2375 2380
 Ph Lys Asp Gln Tyr Lys Asn Asp Asn Ser Asp Asp Asp Asn Val Arg
 2385 2390 2395 2400
 Ser Phe Leu Glu Thr Leu Ile Pro Gln Ile Thr Asp Ala Asn Ala Lys
 2405 2410 2415
 Asn Lys Val Ile Lys Leu Ser Lys Phe Gly Asn Ser Cys Gly Cys Ser
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 2435 2440 2445
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 2450 2455 2460
 Lys His His Gln Thr Ser Asp Thr Glu Cys Ser Asp Thr Pro Gln Pro
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 Gln Thr Leu Glu Asp Glu Thr Leu Asp Asp Asp Ile Glu Thr Glu Glu
 2485 2490 2495
 Ala Lys Lys Asn Met Met Pro Lys Ile Cys Glu Asn Val Leu Lys Thr
 2500 2505 2510
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 2515 2520 2525
 Glu Pro Ala Ala Thr Asp Ser Gly Lys Glu Thr Pro Glu Gln Thr Pro
 2530 2535 2540
 Val Leu Lys Pro Glu Glu Glu Ala Val Pro Glu Pro Pro Pro Pro
 2545 2550 2555 2560
 Pro Gln Glu Lys Ala Pro Ala Pro Ile Pro Gln Pro Gln Pro Pro Thr
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 Pro Pro Thr Gln Leu Leu Asp Asn Pro His Val Leu Thr Ala Leu Val
 2580 2585 2590
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 2595 2600 2605
 Tyr Phe Tyr Leu Lys Val Asn Gly Ser Ile Tyr Met Gly Met Trp Met
 2610 2615 2620
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 2625 2630 2635 2640
 Cys Val Leu Trp Ile Cys Ile Cys Asp Tyr Val Trp Ile Tyr Ile Tyr
 2645 2650 2655
 Ile Tyr Ile Cys Leu Cys Ile Cys Val Phe Gly Tyr Ile Tyr Val Tyr
 2660 2665 2670
 Val Tyr Asp Phe Leu Tyr Met Tyr Leu Trp Val Lys Asp Ile Tyr Ile
 2675 2680 2685
 Trp Met Tyr Leu Tyr Val Phe Tyr Ile Tyr Ile Leu Tyr Ile Cys Ile
 2690 2695 2700
 Tyr Il Lys Lys Glu Ile
 2705 2710

WHAT IS CLAIMED IS:

1. A composition comprising an isolated DABP binding domain polypeptide.

2. The composition of claim 1, wherein the DABP binding domain polypeptide comprises between about 200 and about 300 amino acid residues.

3. The composition of claim 1, wherein the DABP binding domain polypeptide has residues 1 to about 325 of the amino acid sequence of SEQ ID No. 2.

4. The composition of claim 1, wherein the DABP binding region polypeptide is recombinantly produced.

5. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* merozoites in an organism.

6. The composition of claim 5, further comprising an isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* merozoites in an organism.

7. A composition comprising an isolated SABP binding domain polypeptide.

8. The composition of claim 7, wherein the SABP binding domain polypeptide comprises between about 200 and about 600 amino acid residues.

9. The composition of claim 7, wherein the SABP binding domain polypeptide has residues 1 to about 616 of the amino acid sequence of SEQ ID No. 4.

10. The composition of claim 7, wherein the SABP binding region polypeptide is recombinantly produced.

11. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and an isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* merozoites in an organism.

12. The composition of claim 11, further comprising an isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* merozoites in an organism.

13. A composition comprising an isolated polynucleotide which encodes a DABP binding domain polypeptide.

14. The composition of claim 13, wherein the polynucleotide encodes a DABP binding domain polypeptide having residues 1 to about 325 of the amino acid sequence of SEQ ID No. 2.

15. A recombinant cell comprising the polynucleotide of claim 13.

16. A composition comprising an isolated polynucleotide which encodes a SABP binding domain polypeptide.

17. The composition of claim 16, wherein the polynucleotide encodes a SABP binding domain polypeptide having residues 1 to about 616 of the amino acid sequence of SEQ ID No. 4.

18. A recombinant cell comprising the polynucleotide of claim 16.

19. A method of inducing a protective immune response to *Plasmodium* merozoites in a patient, the method comprising administration to the patient of an immunologically effective amount of a pharmaceutical composition comprising a pharmaceutically acceptable carrier and an isolated polypeptide selected from the group consisting of a DABP binding domain polypeptide, an SABP binding domain polypeptide and a combination thereof.

20. The method of claim 19, wherein the patient is a human.

21. A composition comprising a nucleotide sequence of the *EBL* gene family.

22. A composition comprising a nucleotide sequence of the *eb1-e1* gene.

23. The composition of claim 22, wherein the *eb1-e1* gene has the nucleotide sequence of SEQ ID No. 5.

24. A composition comprising the nucleotide sequence of the *E31a* gene.

25. The composition of claim 24, wherein the *E31a* gene has the nucleotide sequence of SEQ ID No. 7.

26. A composition comprising the nucleotide sequence of the *Proj3* gene.

27. The composition of claim 26, wherein the *Proj3* gene has the nucleotide sequence of SEQ ID No. 11.

28. A composition comprising the nucleotide sequence of the *EBL-e2* gene.

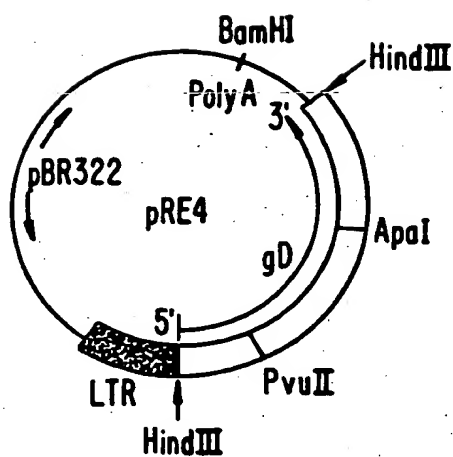
29. The composition of claim 28, wherein the *EBL-e2* gene has the nucleotide sequence of SEQ ID No. 9.

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FAMILY 1	DABP SABP F1 SABP F2 EBL-e1	C-X12 -C-X5--VCIPDRRYOLCMKEL-X47 - DFCKDIRWSLGDGFDIIMGTOMEGIGYSK-X11 - C-X10 -C-X9--VCIPDRRIQLCIVNL-X36 - KFCNDLKNSELDYGHLANGNMDFGGYST-X17 - C-X13 -C-X10 -VCYPPRRQELCLGNI-X36 - EVCKIINKTEADIRDIIGGTDYWNDSLNR-X15 - C-X12 -C-X11 -VCGPPRRQQLCLGYI-X36 - KICNAILGSADIGDIVRGLDVWRDINTN-X17 -
FAMILY 2	EBL-e2 PROJ3 F1 PROJ3 F2 PROJ3 F3 E31a	-----ACAPYRRRLHLCYNL-X43 - OLCTVLARSFADIGDIVRGKOLYLYGDNK-X37 - C-X15 -C-X15 -ACAPYRRRLHVCQNL-X45 - QICTMLARSFADIGDIVRGRLYLGNPQE-X30 - C-X17 -C-X31 -VFLPPRRHEMCTSNL-X55 - AMCRAVRYSFADLGDIIRGRDNWDEKSS-X32 - C-X10 -C-X10 -ACMPRRRQKLCLYYI-X52 - QFLRSMMYTEGDYRDICLNTDISKKQNDV-X15 - C-X10 -C-X11 -ACIPPRRQKLCLYL-X51 - DFKRQMFYTFADYRDICLGTDISKKOTS-X15 -
FAMILY 1 CONT'D	DABP SABP F1 SABP F2 EBL-e1	TDEKAQORRKQWNESKAOIWTAMNYSV-X11 -C-X8 --ePOIYRWIREWGRDYVSELPTEVOKLKEC--X11 --C-X1-- SEHKIKNFREWNEFREKLWEAMLSEH-X6 --C-X6 --eLQITOWIKEWGHEFLERDNRSKLPKSKC--X8 --C-X0-- NKKNDKLFROEWVKVKKDVWNVISWVF-X5 --C-X7 --IPQFRWFSEWGDYCDQDKMIETLKVEC--X4 --C-X1-- KKQNDNERNKWWEKORNLWSSWVKHI-X5 --C-X8 --IPQFLRWLKEWGEFCEENGTEV KoleKIC--X4 --C-X1--
FAMILY 2 CONT'D	EBL-e2 PROJ3 F1 PROJ3 F2 PROJ3 F3 E31a	KGGOFFOLREDWTSNRETIVWKAICH-X11 -C-X23 -VPOYLRWFEEWAEDFCRKKKKKLENLOKOC--X6 --C-X15-- NDPEFFKLRDQWWTANRETIVWKAITCNA-X9 --C-X23 -VPOYLRWFEEWAEDFCRKKKKKKI KDVKRNK--X12 --C-X22-- KKPAYKKLRADWWEANRHQVWRANKCAT-X4 --C-X8 --IPORLRWMTWAEWYCKAOSOEYDKLKKIC--X11 --C-X6-- SKSPSGLSRQEWKTNCPETWKGMLCAL-X37 -----KPOFLRWNIWEGEEFCAEROKKENIKDAC--X8 --C-X3-- KISNSIRYKRSWWTETNGPVIWEGMLCAL-X42 -----RPOFLRWLTWEGENFCKEOKKEYKVLLAKC--X11 --C-X3--
FAMILY 1 CONT'D	DABP SABP F1 SABP F2 EBL-e1	VPPCQACKSYYO WITRKN-X56 -----CX--C EKECIDPCMRYRD WIRSKF-X41 -C-X7 -----CX--C DDNCKSKGNSYKE WISKKK-X36 -C-X20 -----CXX-C EKKCKNACSSYK WIKERN-X38 -C-X19 -----CXX-C
FAMILY 2 CONT'D	EBL-e2 PROJ3 F1 PROJ3 F2 PROJ3 F3 E31a	CTNCSVWGRMYET WIDNOKK-X68 -C-X30 -----CXX-C CISCLYACNPYVD WINNOKE-X69 -C-X40 -----CXX-C CGKCKAACDKYKEEIEKWNEQWRK-X73 -C-X6 -C-X30 -CXX-C KHCRCNOACRAYOE YVENKKF-X43 -C-X4 -----CX--C CVACKDCKOYNS WIGIWD-X42 -C-X8 -----CXXC

FIG. 1.

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**FIG. 2.**

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- I. UNIEBP5 AND 5A: P R R Q K/E L C
 UNIEBP5, FOR A+T BIASED CODON USAGE:
 CC(A/G)-AG(G/A)-AG(G/A)-CAA-(G/A)AA-(C/T)TA-TG
 UNIEBP5A, FOR G+C BIASED CODON USAGE:
 CC(C/G)-(C/A)G(C/G)-(C/A)G(C/G)-CAG-CAG-(C/T)T(C/G)-TG
- II. UNIEBP5 B AND C: F A D VY G/R D I
 UNIEBP5B, FOR A+T BIASED CODON USAGE:
 TTT-GC(A/T)-GAT-(A/T)(A/T)(A/T)-(G/C)G(A/T)-GAT-AT
 UNIEBP5C, FOR G+C BIASED CODON USAGE:
 TTC-GC(G/C)-GAT-(A/TXA/T)C-(G/C)G(G/C)-GAC-AT
- III. UNIEBP3 AND 3A: P Q F L/F R W
 UNIEBP3, FOR A+T BIASED CODON USAGE:
 CCA-(A/T)C(T/G)-(T/G)A(A/G)-(A/G)AA-TTG-(A/T)GG
 UNIEBP3A, FOR G+C BIASED CODON USAGE:
 CCA-(C/G)C(G/T)-G(A/T)A-GA(A/T)-CTG-(C/G)GG
- IV. UNIEBP3 B AND C: E W G D/E D/E Y/F C
 UNIEBP3B, FOR A+T BIASED CODON USAGE:
 CA-A(A/T)A-(A/T)TC-(A/T)TC-(A/T)CC-CCA-TTC
 UNIEBP3C, FOR G+C BIASED CODON USAGE:
 CA-G(A/T)A-(G/C)TC-(G/C)TC-(G/C)CC-CCA-CTC G-C BIASED

FIG. 3.